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(54) **Diagnostic reagent for hepatitis C.**

(57) A diagnostic reagent for hepatitis C, which detects an antibody induced by infection of hepatitis C virus, characterized in that said diagnostic reagent comprises the second envelope protein or the first non-structural protein which is encoded by the gene of hepatitis C virus and has a sugar chain. This invention also provide a method for detecting an anti-hepatitis C virus antibody. The use of the diagnostic reagent for hepatitis C according to the present invention makes highly sensitive diagnosis of hepatitis C possible.

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BACKGROUND OF THE INVENTION

This invention relates to a diagnostic reagent for hepatitis C comprising an antigen protein translated from a genome of hepatitis C virus. More specifically, this invention relates to a diagnostic reagent for detecting an antibody against hepatitis C virus (hereinafter referred to as "HCV"), which comprises a protein encoded by a gene of HCV, wherein said protein is identified as a glycoprotein called the second envelope protein or the first non-structural protein (hereinafter referred to as "E2/NS1").

The first successful cloning of human hepatitis virus which had been called non-A, non-B hepatitis virus was accomplished in 1988 by Chiron Co., Ltd. U.S.A and the hepatitis virus was designated HCV. Further, Chiron Co., Ltd. succeeded in expressing in a yeast a fused protein which comprises at the C-terminal the polypeptide corresponding to the region having 363 amino acid residues from the third nonstructural protein (NS3) to the fourth non-structural protein (NS4) both of which are portions of nonstructural proteins of HCV and at the N-terminal human superoxide dismutase (European unexamined patent publication No. 318216) and, using this recombinant antigen, developed a diagnostic reagent for hepatitis C (Science, 244, 359-362, 362-364, (1989)).

In Japan, the Japanese Red Cross Society has been using the diagnostic reagent in the screening of blood provided by donors, which is known as "C100-3 antibody test", in order to avoid post-transfusion hepatitis since the end of 1989. However, since not all samples are effectively screened only by C100-3 antibody test, post-transfusion hepatitis is not completely avoided.

Subsequently, further investigation of HCV genomes derived from the serum of a Japanese patient by the cloning technique revealed that HCV prevailed in Japan is similar to HCV obtained by Chiron Co., Ltd. but a different strain (Protein, Nucleic acid and Enzyme, 36, 1679-1691, (1991)). In addition, the use of the core protein (C) region of the structural protein, the third non-structural protein (NS3) region, the fifth non-structural protein region and the like have been proposed as more effective diagnostic reagents than C100-3 (Lancet, 337, 317-319, 1991 and Japanese unexamined patent publication (hereinafter referred to as "J. P. KOKAI") No. Hei 3-103180).

The C100-3 antibody test system has a disadvantage that the detection rate and the sensitivity are low as mentioned above. Although proteins derived from C, NS3 and NS5 regions have been proposed as more effective antigens for detection than C100-3, any satisfactory results have not yet been reported. Therefore, there is a need for a diagnostic reagent and a diagnostic method for hepatitis C, having a higher detection rate and sensitivity.

SUMMARY OF THE INVENTION

The inventors have conducted various investigations to obtain a diagnostic reagent for hepatitis C, having a higher detection rate and sensitivity. As a result, they have found that E2/NS1 protein having a sugar chain, which is obtained by expressing cDNA of E2/NS1 region in animal cells reacts with the serum of the patient of hepatitis C with a high rate in a fluorescent antibody test and accomplished the goals of the present invention. The high reaction rate of E2/NS1 region with the serum of the patient of hepatitis C was unexpected because the protein derived from E2/NS1 region is susceptible to the mutation of an amino acid sequence and, therefore, the protein expressed in *E. coli* has been considered to react with the serum of the patient of hepatitis C with a lower rate comparing with the proteins derived from the other regions of HCV and it has not been expected to use the protein for a diagnostic reagent.

The present invention provides a diagnostic reagent for hepatitis C, which detects an antibody induced by infection of hepatitis C virus, characterised in that said diagnostic reagent comprises the second envelope protein or the first non-structural protein which is encoded by the genome of hepatitis C virus and has a sugar chain.

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 shows the steps of constructing DNA fragment 1325SK containing the base sequence of clone J1-1325.

Fig. 2 shows the steps of constructing plasmid pSR316EP.

Fig. 3 shows the steps of constructing plasmid pSRNot.

Fig. 4 shows the steps of constructing expression vector paSR1325X-3 having a DNA fragment coding for E2/NS1 protein.

Fig. 5 shows the steps of constructing plasmid pHLp1.

Fig. 6 shows the steps of constructing expression vector mulcos pHL16SR1325 having 16 DNA fragments coding for E2/NS1 protein.

DETAILED EXPLANATION OF THE INVENTION

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E2/NS1 protein of the present invention is a protein derived from the region called the second envelope protein or the first nonstructural protein, which is encoded by the genome of HCV. Examples of the proteins are illustrated in SEQUENCE ID Nos.1-12 in SUQUENCE LISTING. Proteins obtained from such proteins by deleting, inserting, modifying or adding a part of amino acids are encompassed in the scope of the present invention provided that they maintain the reactivity with the serum of the patient of hepatitis C.

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(1) Method of preparing clones of cDNA derived from the serum of the patient of hepatitis C, which are shown in SEQUENCE ID Nos. 1-3 of SEQUENCE LISTING and determining the base sequence thereof

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Genes or DNA fragments coding for novel polypeptides, which are shown in SEQUENCE ID Nos. 1-3 of SEQUENCE LISTING can be prepared, for example, by a method described below.

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Since there exists a trace of HCV in the serum and the genome of HCV is expected to be RNA, it was expected that cloning by Okayama-Berg method or Gubler-Hoffman method of the prior art would be attended by difficulties and, therefore, the following method was conducted to ensure the cloning of the gene susceptible to mutation from a trace of the serum.

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The nucleic acid is extracted from the serum of the patient of hepatitis C as described in Example 1 later. Generally, it is preferred to use the serum having an OD value of 3.5 or more measured by a test kit of Ortho Inc. However, the present invention is not limited to the use of the serum having such an OD value. The serum is preferably mixed with transfer RNA (tRNA) as a carrier of virus RNA. The carrier is not limited to tRNA. Any polyribonucleoside can be used as carriers. If tRNA is used, there is an advantage that it can be rapidly confirmed by electrophoresis whether there is a required amount of tRNA having an intact length. By this confirmation, it can also be confirmed whether virus RNA degrades after being mixed with tRNA as a carrier of virus RNA. As a technique of cloning cDNA from the nucleic acid, it is preferred to use polymerase chain reaction method developed by Saiki et al. (PCR method, Nature, 324, 126, (1986)). First of all, a reverse transcriptase is reacted using virus RNA as a template. In the reaction, any commercially available random primers or synthesized DNA having a base sequence similar to that of primer AS1 which is shown below may be used as a primer.

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5'

3'

AS1:GCTATCAGCAGCATCATCCA SEQUENCE ID No.13

A few bases at the 5' end of these sequences may be changed to other bases. Preferably, a few bases within 10 bases from the 5' end and more preferably, a few bases within 5 bases from the 5' end may be changed to other bases. In addition, 4-5 bases, preferably a few bases may be deleted from the sequences at the 5' end of these sequences. Furthermore, any 8-12 bases, preferably 5-6 bases, more preferably a few bases, may be added to the sequences at the 5' end of these sequences.

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PCR method is specifically carried out under the conditions described in Example 1. PCR method is carried out as described in Example 1 using the first complementary DNA (1st cDNA) thus obtained as a template to prepare a desired DNA fragment. The conditions of PCR method are suitably selected depending on the circumstances. Representative examples of sense primers include the following one:

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5'

3'

S1:CAGITAITCCGGATCCCICAAG SEQUENCE ID No.14

"I" appearing in the sequenc means inosine. A few bases at the 5' end of these sequences may be changed to other bas s. Preferably, a few bases within 10 bases, more preferably, within 5 bases from the 5' end may b changed to oth r bases. In addition, 4-5 bases, preferably a few bases may be deleted from the sequences at the 5' end of these sequences. Furthermore, any 8-12 bases, preferably 5-6 bases, more

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preferably a few bases may be added to the sequences at the 5' end of these sequences.

The DNA fragment thus obtained is inserted at one of cloning sites such as Sma I site of a cloning vector such as pUC19 according to conventional technique. Using a plasmid having this DNA fragment, the base sequences of at least 3 clones are determined independently regarding the both strands. The determination of the base sequences can be easily carried out by a dideoxy method using, for example, 7-deaza sequence kit available from Takara Shuzo Co., Ltd. or fluorescence sequencer GENESIS 2000 system available from Du Pont according to the protocol thereof. When the DNA fragment has a site which is considered difficult to determine the base sequence or has more than about 180 base pairs, a subcloning may be carried out according to conventional technique. SEQUENCE ID Nos.1-3 of SEQUENCE LISTING show the amino acid sequences of the proteins assumed from the base sequences of the DNA fragments thus determined.

Clone J1-1325 (SEQUENCE ID No.1), clone N27, clone N19, H19 and Y19 (SEQUENCE ID No. 3) were prepared with the serums of different patients. Clone MX24 (SEQUENCE ID No.3) was prepared with a pool of the serums of the patients of hepatitis C. The clones shown in SEQUENCE ID Nos.1-3, which were prepared using a combination of primer S1 with primer AS1 correspond to the same region in the gene of HCV.

Antigen proteins derived from E2/NS1 protein regions shown in SEQUENCE ID Nos.4-12 of SEQUENCE LISTING can also be used in the present invention.

The antigen protein of SEQUENCE ID No.4 can be obtained by expressing cDNA described in Journal of Virology, 65, 1105-1113, (1991). The antigen protein of SEQUENCE ID No.5 can be obtained by expressing cDNA described in Proceedings of the National Academy of Sciences of the USA, 87, 9524-9528, (1990). The antigen protein of SEQUENCE ID No.6 can be obtained by expressing cDNA described in The fiftieth general meeting of Japanese Cancer Society, 379, (1991). The antigen protein of SEQUENCE ID No.7 can be obtained by expressing cDNA described in European Patent No.0,388,232 (1990). The antigen proteins of SEQUENCE ID Nos.8 and 9 can be obtained by expressing cDNAs described in Proceedings of the National Academy of Sciences of the USA, 88, 3392-3396, (1991). The antigen proteins of SEQUENCE ID Nos.10 and 11 can be obtained by expressing cDNAs described in Japanese Journal of Experimental Medicine, 60, 167-177, (1990). The antigen protein of SEQUENCE ID No.12 can be obtained by expressing cDNA described in Biochemical and Biophysical Research Communications, 175, 220-228, (1991). The sequences shown in SEQUENCE ID Nos.4-12 correspond to the same region as that of the sequences shown in SEQUENCE ID Nos.1-3. (2) Expression of polypeptides encoded by the clones prepared in step (1)

In order to produce E2/NS1 protein, it is necessary to select an appropriate host-vector system which is able to stably express the protein. Further, it is required that the expressed E2/NS1 protein has the same level of biological activity, that is, antigenicity as that of HCV. Considering that natural E2/NS1 protein is expected to be a glycoprotein and that E2/NS1 protein contains many cysteine residues and the positions of the thiol bonds between the cysteine residues and the higher-order structure of the protein are important to maintain the activity, it is desired to express the protein in such an animal cell host as CHO cell, COS cell, mouse L cell, mouse C127 cell and mouse FM3A cell, preferably CHO cell. When these cells are used as hosts, it is expected that processed E2/NS1 protein is produced by introducing E2/NS1 gene having a signal-like sequence of from the 32 position to the 44 position of the amino acid sequences shown in SEQUENCE ID Nos.1-12 into the cell. Expression plasmids for these animal host cells can be constructed as follows:

As promoters in the animal cells, one can use the active-type promoter of adenovirus E1A gene (Biochemical Experiment Lecture, second series, Vol. 1, Techniques for gene investigations II, 189-190 (1986)), the early promoter of SV40, the late promoter of SV40, the promoter of apolipoprotein E gene and SR α promoter (Molecular and Cellular Biology, 8, 466-472, (1988)), preferably the promoter of SV40 and SR α promoter.

A DNA fragment of a gene coding for E2/NS1 protein containing the signal-like sequence is inserted downstream of the promoter in a direction of the transcription. When the expression vector of E2/NS1 protein is constructed, a ligated gene fragment of at least two gene fragments coding for E2/NS1 protein may be inserted downstream of the promoter. At least two units of DNA fragments ligated upstream of the 5' end of the DNA fragment of the gene coding for E2/NS1 protein with such a promoter as that of SV40 may be ligated together in the same direction of the transcription and then inserted in the vector. Polyadenylation sequence is required to be present downstream of the gene coding for E2/NS1 protein. For example, at least one of polyadenylation sequences derived from SV40 gene, β -globin gene or metallothionein gene is required to be present downstream of the gene coding for E2/NS1 protein. When at least two of the DNA fragments containing the gene coding for E2/NS1 protein ligated to the promoter are

ligated, the polyadenylation sequence may be present at each 3' end of the gene coding for E2/NS1 protein.

In transforming an animal cell such as CHO cell with this expression vector, the use of a selective marker is desired. Examples of the selective markers include DHFR gene expressing methotrexate resistance (Journal of Molecular Biology, 159, 601, (1982)), Neo gene expressing antibiotic G-418 resistance (Journal of Molecular Applied Genetics, 1, 327, (1982)), Ecogpt gene derived from E. coli, expressing mycophenol acid resistance (Proceedings of the National Academy of Sciences of the USA, 78, 2072, (1981)), hph gene expressing antibiotic hygromycin resistance (Molecular and Cellular Biology, 5, 410, (1985)) and the like. A promoter such as the aforementioned promoter derived from SV40 and the promoter of TK gene of Herpes virus is inserted upstream of the 5' end of each drug resistance gene. The aforementioned polyadenylation sequence are contained downstream of the 3' end of each drug resistance gene. When such a drug resistance gene is inserted in the expression vector of E2/NS1 protein, it may be inserted downstream of the polyadenylated site in the gene coding for E2/NS1 protein in a right direction or a reverse direction. These expression vectors do not require any co-transfection with another plasmid containing a selective marker gene in preparing a transfect.

In the case where such a selective marker gene is not inserted in the expression vector of E2/NS1 protein, a vector having a selective marker of the transfect, such as pSV2neo (Journal of Molecular Applied Genetics, 1, 327, (1982)), pMBG (Nature, 294, 228, (1981)), pSV2gpt (Proceedings of the National Academy of Sciences of the USA, 78, 2072, (1981)), pAd-D26-1 (Journal of Molecular Biology, 159, 601, (1982)) and the like may be used together with the expression vector of E2/NS1 protein to conduct co-transfection. The transfect can be easily selected by gene expression of the selective marker gene.

Examples of methods of introducing the expression vector into the animal cell include calcium phosphate method (Virology, 52, 456, (1973)) and electroporation method (Journal of Membrane Biology, 10, 279, (1972)). Calcium phosphate method is used in general.

The transfected animal cell can be cultured by a float culture or an adherent culture in the conventional manner. The cultivation can be conducted in a medium such as MEM, Ham, F-12 and the like in the presence of 5-10 % of serum or a suitable amount of insulin, dexamethasone and transferrin or in the absence of serum. The animal cell expressing E2/NS1 protein can be detected by fluorescent antibody technique using the serum of the patient according to the conventional method. The cloning is carried out by limiting dilution according to the conventional method to establish a cell line stably producing E2/NS1 protein.

E2/NS1 protein derived from HCV gene, thus obtained can be used as HCV antigen which reacts immunologically with the serum containing HCV antibody and therefore, is useful for the confirmation or the detection of the presence of Anti-HCV antibody in samples including blood or serum. Examples of the immunoassays include RIA (radioimmunoassay), ELISA (enzyme-linked immunoadsorbent assay), fluorescent antibody technique, agglutination reaction including latex fixation, immuno precipitation and the like. In the detection, a labelled antibody is usually used. A labelling substance such as a fluorescent substance, a chemoluminescent substance, a radioactive substance, a dyeing substance and the like can be used. Accordingly, using the above E2/NS1 protein derived from HCV gene as an antigen, the diagnostic reagent for hepatitis C according to the present invention can be prepared.

The reagent containing the protein having a sugar chain, which is derived from E2/NS1 region according to the present invention makes the confirmation or the detection of the presence of anti-HCV antibody in samples including blood or serum possible. The use of the reagent according to the present invention makes highly sensitive diagnosis of hepatitis C possible.

The present invention will be explained in more detail with reference to the following non-limiting examples.

Example 1

(1) Extraction of the nucleic acid from the serum of the patient of hepatitis C

Twenty-five milliliters of a Tris buffer (50 mM Tris-HCl (pH 8.0), 1 mM EDTA and 100 mM NaCl) were added to 10 ml of the serum of the patient of hepatitis C, which showed at least 3.5 of an OD value by a HCV EIA kit available from Ortho Inc. After being mixed, the mixture was centrifuged at 20,000 x g at 20 °C for 20 minutes. The obtained supernatant was centrifuged at 100,000 x g at 20 °C for additional 5 hours. One point five milliliters of a Proteinase K solution (1% sodium dodecyl sulfate, 10 mM EDTA, 10mM Tris-HCl (pH 7.5), 2 mg/ml Proteinase K (available from Pharmacia Co.) and 6.6 µg of a yeast tRNA mixture) were added to the precipitate. After the precipitate was dissolved in the Proteinase K solution, the obtained

solution was maintained at 45°C for 90 minutes. The mixture was subjected at least four times to a phenol/chloroform treatment which comprises the steps of adding an equivalent amount of phenol/chloroform, violently agitating and then centrifuging the mixture to collect an aqueous phase containing a nucleic acid. Then, a chloroform treatment was carried out at least 2 times. To the obtained aqueous phase, one-tenth amount of 3M sodium acetate or an equivalent amount of 4M ammonium acetate, and 2.5-fold volume of ethanol were added and the mixture was left to stand at -20 °C overnight or -80 °C for at least 15 minutes. The mixture was centrifuged at 35,000 rpm for 4 hours by a SW41Ti rotor (available from Beckmann Co.) to collect a nucleic acid as a precipitate.

10 (2) Synthesis of cDNA

(2-1) Synthesis of an RNA sample

After the nucleic acid obtained in step (1) was dried, 30 µl of water and 10 µl of ribonuclease inhibitor (100 units/ µl, available from Takara Shuzo Co., Ltd.) were added thereto to dissolve the nucleic acid. The following synthesis of cDNA was carried out using the obtained nucleic acid solution.

(2-2) Synthesis of cDNA using an anti-sense primer

To 2 µl of the aqueous solution of the nucleic acid prepared in step (2-1), 1 µl of an anti-sense primer (synthesized DNA primer AS1; 15 pmoles/ µl), 2 µl of 10xRT buffer (100mM Tris-HCl (pH 8.3) and 500 mM of KCl), 4 µl of 25 mM MgCl₂, 8 µl of 2.5 mM 4dNTP and 1 µl of water were added and the mixture was maintained at 65°C for 5 minutes and at room temperature for 5 minutes. Subsequently, 1 µl of 25 units of a reverse transcriptase (available from Life Science Co.) and 1 µl of a ribonuclease inhibitor (100 units/ µl, available from Takara Shuzo Co., Ltd.) were added to the mixture and then the resulting mixture was maintained at 37 °C for 20 minutes, then at 42 °C for 30 minutes and finally at 95 °C for 2 minutes. Immediately thereafter, the mixture was cooled to 0°C (Synthesis of complementary DNA). The DNA having a specific sequence was amplified using 10 µl of the DNA sample according to Saiki's method (Nature, 324, 126, (1986)), so-called PCR method as follows:

Water was added to a mixture of 10 µl of the above DNA sample, 10 µl of 10xPCR buffer (100 mM of Tris-HCl (pH 8.3), 500 mM of KCl, 15 mM of MgCl₂, and 1 % of gelatin), 8 µl of 2.5 mM 4dNTP, 2 µl of the synthesized DNA primer used in the synthesis of the complementary DNA (150 pmoles/ µl), 3 µl of a synthesized DNA primer corresponding to the DNA primer (15 pmoles/ µl) (which is complementary to the synthesized DNA primer used in the synthesis of the complementary DNA, i.e., the aforementioned primer S1) to prepare 100 µl of an aqueous solution. After the solution was maintained at 95°C for 5 minutes, it was cooled rapidly to 0°C. One minute after the cooling, the solution was mixed with 0.5 µl of Taq DNA polymerase (7 units/ µl, Trade Name "AmpliTaQ™" available from Takara Shuzo Co., Ltd.) and then mineral oil was layered on the mixture. This sample was incubated on a DNA Thermal Cycler available from Parkin Elmer Cetus Co. at 95 °C for 1 minute, at 40-55 °C for 1 minute, and at 72°C for 1-5 minutes for 25 cycles. After the sample was incubated finally at 72 °C for 7 minutes, the reaction aqueous solution was subjected to a phenol/chloroform treatment and a precipitation treatment with ethanol to obtain amplified DNA fragments. The above precipitation treatment with ethanol was carried out by mixing the aqueous phase with a one-tenth amount of 3 M sodium acetate or an equivalent amount of 4 M ammonium acetate together with a 2.5-fold volume of ethanol, centrifuging the mixture at 15,000 rpm at 4°C for 15 minutes by a rotor having a radius of about 5 cm and drying the precipitate.

(3) Cloning of the amplified DNA fragments and Determination of the base sequences thereof

At least 1 pmole of the DNA fragments obtained by the method described in step (2-2) was treated with T4 DNA polymerase (available from TOYOBO CO.,LTD) to make blunt ends (Molecular Cloning, 1982, Cold Spring Harbor Laboratory Press). After a phosphoric acid group was introduced into the DNA fragment at the 5' end with polynucleotidekinase (available from TOYOBO CO.,LTD) (Molecular Cloning, 1982, Cold Spring Harbor Laboratory Press), the DNA fragment was inserted at Sma I site present in the multicloning sites of pUC19 cloning vector using a ligation kit (available from Takara Shuzo Co., Ltd.).

The vector DNA prepared in the following procedure was used in the ligation in an amount of 5-10 ng. pUC18 cloning vector was cleaved with restriction enzyme Sma I (available from TOYOBO CO.,LTD) and then subjected to a phenol/chloroform treatment and a precipitation treatment with ethanol. Subsequently, this was treated with alkaline phosphatase (available from Boehringer Mannheim) to conduct the

dephosphorylation at the 5' end (Molecular Cloning, 1982, Cold Spring Harbor Laboratory Press), followed by a phenol/chloroform treatment and a precipitation with ethanol. The competent cell of *E. coli* JM109 or DH5 (available from TOYOBO CO., LTD) was transformed with the DNA prepared in the above procedure. The procedure of the transformation was according to the protocol of COMPETENT HIGH prepared by TOYOBO CO., LTD. At least 20 transformants transformed with the pUC18 cloning vector having the DNA fragment obtained by the method described in step (2-2) using the combination of the aforementioned primers were prepared.

Plasmid DNA pUC1325 shown in Fig. 1 was prepared from the obtained transformant in the conventional method and the base sequence of the plasmid was determined by a 7-deaza sequence kit available from Takara Shuzo Co., Ltd. or a fluorescence sequencer GENESIS 2000 system available from Du Pont. Two kinds of synthesized primers, 5'd(GTAAACGACGGCCAGT)3' (SEQUENCE ID No. 15) and 5'd-(CAGGAAACAGCTATGAC) 3' (SEQUENCE ID No. 16) were used to determine a base sequence of the + strand and that of the - strand of the DNA fragment. The DNA fragment had the same base sequence as that shown in SEQUENCE ID No. 1 of SEQUENCE LISTING. The amino acid sequence shown in SEQUENCE ID No. 1 of SEQUENCE LISTING is encoded by the + strand of the gene derived from HCV and inserted in the plasmid of the transformant.

The amino acid sequence encoded by the DNA fragment obtained was compared with the reported sequences of hepatitis C viruses. In step (2-2) of Example 1, three clones were obtained from the serum of one patient. The determination of the base sequence of the clones reveals that the patient carries several kinds of viruses.

(4) Preparation of a plasmid expressing E2/NS1 protein

Figs. 1-6 show a procedure of preparing a plasmid expressing E2/NS1 protein.

(4-1) Preparation of DNA fragment 1325SK

The DNA fragment of clone 1325 contained in plasmid pUC1325 obtained in step (3) was inserted at Sma I site of pUC18 so that the fragment had KpnI site of pUC18 at the 5' end of the + strand of clone 1325 coding for E2/NS1 protein and Bam HI site of pUC18 at the 3' end. After complete digestion with restriction enzyme Hin dIII, the fragment was partially digested with restriction enzyme Bam HI to obtain a DNA fragment which was cleaved not at Bam HI site within the vector but only at another Bam HI site present in clone 1325. The DNA fragment contains from the Bam HI site present at the 5' end to the 3' end of clone 1325 which was the DNA fragment obtained in step (2-2), which was derived from the gene of HCV.

Subsequently, as shown in Fig. 1, the DNA fragment was treated with T4 DNA polymerase to make blunt ends. After being ligated with SpeI linker consisting of the sequence of 5' pGGACTAGTCC 3' (SEQUENCE ID No. 17) (available from New England Biolab Co.), the fragment was cleaved with restriction enzyme Xba I (the Xba I site of the fragment was derived from plasmid pUC18). The following adaptor was ligated to Xba I site at the 3' end to obtain DNA fragment 1325SK.

5' pCTAGAGAATTCGGTAC 3' (SEQUENCE ID No. 18)

3' TCTTAAGCp 5'

(4-2) Construction of plasmid pSRNot

Expression vector pAC316 reported in Journal of Virology, 65, 3015-3021, (1991) was cleaved with restriction enzyme Tth 111I at Tth111I site present at the 3' end of 3' poly A region. T4 DNA polymerase was acted on the cleaved vector to make blunt ends. The fragment between SalI site and Eco RI site of plasmid pmoRH (Fig. 2) reported by Ikeda et al (Gene, 71, 19-27, (1988)) was cut out and T4 DNA polymerase was acted on the fragment to make blunt ends.

As shown in Fig. 2, the DNA fragment derived from pAC316 and the DNA fragment derived from pmoRH were ligated together with Bgl II linker (available from Takara Shuzo Co., Ltd.) to obtain plasmid pSR316EP containing one Bgl II linker and one DNA fragment containing the early promoter of SV40 derived

from pmoRH. As shown in Fig. 3, after plasmid pSR316EP was cleaved with restriction enzymes Hgi AI and Dra III, T4 DNA polymerase was acted on the plasmid to make blunt ends. Then, one Not I linker was introduced in the plasmid to obtain plasmid psRNot (Fig. 3). Namely, Not I linker was prepared by synthesizing DNA having a sequence of 5' AGCGGCCGC 3' and phosphorylating the 5' end by kination

(Molecular Cloning second edition, 11.31-11.44, (1989), Cold Spring Harbor Laboratory Press).
Subsequently, dhfr gene was cut out from plasmid pCHD2L reported by Ikeda et al in Gene, 71, 19-27, (1988) using restriction enzymes Kpn I and Eco RV and Kpn I-EcoRV fragment of plasmid Charomid9-36 described in Proceedings of the National Academy of Sciences of the USA, 83, 8664-8668, (1986) was inserted in the deleted dhfr gene region instead of the KpnI-EcoRV fragment coding for dhfr gene as shown in Fig. 5 to obtain plasmid pChmBp1. The plasmid contains a polylinker derived from plasmid Charomid9-36.

Then, plasmid pAG60 reported by Garapin et al. in Journal of Molecular Biology, 150, 1-14, (1981) was cleaved with restriction enzyme Pvu II to obtain a Pvu II fragment coding for a neomycin gene. After plasmid pChmBp1 was cleaved with restriction enzyme Eco RV and then T4 DNA polymerase was acted to make blunt ends, the fragment obtained was ligated to the Pvu II fragment to obtain plasmid pHLp1 which contained the neomycin gene derived from plasmid pAG60 at the Eco RV site of plasmid pChmBp1 (Fig. 5).

(4-3) Construction of expression vector paSR1325X-3

As shown in Fig. 4, after plasmid pSRNot obtained in step (4-2) was cleaved with restriction enzyme Not I and then with T4 DNA polymerase to make blunt ends, this was cleaved with restriction enzyme Kpn I. The obtained DNA fragment was ligated to DNA fragment 1325SK obtained in step (4-1) to obtain expression vector paSR1325X-3 having only one DNA fragment 1325SK (Fig. 4).

(4-4) Construction of expression vector pHL16SR1325

As shown in Fig. 6, expression vector paSR1325X-3 obtained in step (4-3) was cleaved with restriction enzyme Sfi I to prepare two fragments one of which was an expression unit of clone 1325. The Sfi I sites were present in an initial promoter of SV40. Five μ g of the Sfi I fragment having the expression unit of clone 1325 was ligated to 50 ng of the fragment obtained by cleaving expression vector pHLp1 with restriction enzyme Sfi I in 10 μ l of a reaction solution using a ligation kit available from Takara Shuzo Co., Ltd. according to a protocol for the ligation kit to obtain expression vector pHL16SR1325 (Fig. 6).

The vector had successive sixteen DNA fragments 1325SK having at the Sfi I site of expression vector paSR1325X-3 the expression unit of clone 1325 which was a gene coding for E2/NS1 protein. In the vector, all of the DNA fragments 1325SK were inserted downstream of SV40 promoter of expression vector paSR1325X-3 in a direction of transcription.

(5) Obtaining a cell line constantly expressing E2/NS1 protein

Expression vector pHL16SR1325 prepared in step (4) was recovered from the recombinant E.coli DH1 strain, purified according to the conventional technique described in Molecular Cloning second edition, 1989, Cold Spring Harbor Laboratory Press to obtain a large amount of the expression plasmid DNA. CHO cells were transfected with the plasmid DNA according to the method described in Ausubel et al. (Current Protocols in Molecular Biology, Greene Publishing Associates and Wiley-Interscience, Chapter 9.1.1-9.1.4, (1987)) as follows:

CHO cells were cultured in Ham F-12 medium containing 10 % of fetal calf serum (FCS) in a plate having a diameter of 6 cm until the cells were in semiconfluent condition. Then, the medium was removed from the plate and a DNA solution was dropwise added thereto. The DNA solution was previously prepared by the following procedure.

Three hundred μ l of 2xHEBS solution (2xHEBS solution; 1.6 % sodium chloride, 0.074 % potassium chloride, 0.05 % $\text{Na}_2\text{HPO}_4 \cdot 12\text{H}_2\text{O}$, 0.2 % dextrose and 1 % HEPES (pH 7.05)) were mixed with 10 μ g of the plasmid DNA in each plate and sterilized water was added to the mixture to prepare a solution of 570 μ l. The solution was charged in an Eppendorf centrifuge tube. The DNA solution was violently agitated by a Vortex mixer for 1-2 seconds while adding 30 μ l of 2.5 M calcium chloride solution thereto. The DNA solution was agitated by a Vortex mixer at about 10-minute intervals during being left to stand for 30 minutes. The obtained DNA solution was added to the aforementioned CHO cells and the CHO cells were left to stand at room temperature for 30 minutes. Then, 5 ml of Ham F-12 medium containing 10 % of FCS

available from GIBCO Co. were added to the plate and the culture was incubated at 37 °C under air containing 5 % carbon dioxide for 4-5 hours. Subsequently, the medium was removed from the plate and the cells were washed with 5 ml of a 1xTBS ++ solution (1xTBS ++ solution ; 25 mM Tris-HCl (pH 7.5), 140 mM sodium chloride, 5mM potassium chloride, 0.6 mM disodium hydrogen phosphate, 0.08 mM calcium chloride and 0.08 mM magnesium chloride). After the 1xTBS ++ solution was removed, 5 ml of a 1xTBS ++ solution containing 20 % of glycerol was added to the cells and the culture was left to stand at room temperature for 1-2 minutes. After the supernatant was removed from the plate, the cells were washed again with 5 ml of a 1xTBS ++ solution and cultured in 5 ml of fresh Ham F-12 medium containing 10 % of FCS in the plate at 37 °C under air containing 5 % carbon dioxide for 48 hours. Then, the medium was removed and the cells were washed with 5 ml of a 1xTBS ++ solution. The cells were treated with a trypsin-EDTA solution (available from Sigma Co.) and left to stand at room temperature for 30 seconds. Five minutes after the trypsin-EDTA solution was removed, the cells attached to the wall of the plate were peeled adding 5 ml of Ham F-12 medium containing 10 % of FCS. The cells cultured in one plate having a diameter of 5 cm were divided in ten plates having a diameter of 9 cm and cultured in the plates containing drug G418 (G418 sulfate (GENETICIN) available from GIBCO Co.) in a concentration of 600 µg/ml. Ten days after the cultivation, grown cells having G418 resistance were isolated and cultured for about 7 days in 1 ml of Ham F-12 medium containing 10 % of FCS in a 24 well titer plate each well of which has an area of about 3.1 cm².

A part of the cells were cultured on slide glass (Lab-Tek Chamber Slides, Nunc4808 available from Japan Inter Med Co.) overnight. After being rinsed with phosphate buffered saline (PBS), the slide glass was immersed in cold acetone-methanol (1:1) solution and maintained at -20 °C for 15 minutes to fix the cells. The cells fixed on the slide glass were reacted with the serum of the patient of hepatitis C 20-fold diluted with PBS at 37 °C for 30 minutes. Then, the slide glass was washed three times with PBS for 5 minutes and reacted with FITC-labelled rabbit anti-human IgG (available from Daco Japan Co.) 50-fold diluted with PBS at 37 °C for 30 minutes. The slide glass was washed three times with PBS for 5 minutes and dried by putting the slide glass between two pieces of filter paper. After the slide glass was sealed with glycerin, the cells on the slide glass were observed under a fluorescence microscope. Screening positive cells as described above, successive three times of limiting dilution were carried out to establish cell line 13L20 constantly producing E2/NS1 protein.

(6) Study of the reactivity of 13L20 cells with the serum of the patient of hepatitis C

After 13L20 cells established in step (5) were cultured on Lab-Tek Chamber Slides (Lab-Tek Chamber Slides, Nunc4808 available from Japan Inter Med Co.) overnight and then fixed with a cold acetone-methanol solution, the fixed cells were reacted with 59 serum samples of the patients of hepatitis C. Then, the cells were washed as described above and reacted with the secondary antibody. The observation under a fluorescence microscope revealed that 53 samples were positive. Among the 59 serum samples, 6 samples were judged to be positive using CHO cells constantly producing the first envelope region of HCV.

Example 2

Using as a template the DNA fragment described in Example 11 (3) of the specification of European Patent Application No. 92109812.5 filed on June 11, 1992 (TITLE OF THE INVENTION "Gene or DNA fragments derived from hepatitis C virus, polypeptides encoded thereby, and method of producing thereof"), PCR reaction was carried out in the same manner as that of Example 1 using the same primer to obtain a DNA fragment corresponding the same region as that of clone J1-1325 shown in SEQUENCE ID No. 1 of SEQUENCE LISTING. The region was a DNA fragment encoding for E2/NS1 protein like clone J1-1325. For example, using as a template the DNA fragment clone N27MX24A-1 having a base sequence shown in SEQUENCE ID No. 31 of SEQUENCE LISTING described in the specification of the aforementioned European Patent Application filed on June 11, 1992, plasmid pUCN27MX24A-2 was obtained. The base sequence of the DNA fragment coding for E2/NS1 protein, which was cloned in the plasmid is shown in SEQUENCE ID No. 2 of SEQUENCE LISTING. In addition, MK2724A2 cell line constantly producing E2/NS1 protein was stabilised by the same procedure as that described in steps (4) and (5) of Example 1. The reactivity of the same samples as Example 1 with the cell line was estimated by the same method as that described in step (6) of Example 1. Results similar to those obtained in step (6) of Example 1 were obtained.

SEQUENCE LISTING

(2) INFORMATION FOR SEQ ID NO:1:

5

(i) SEQUENCE CHARACTERISTICS:

10

(A) LENGTH: 1207 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

15

(D) TOPOLOGY: linear

20

(iv) ANTI-SENSE: No

(vi) ORIGINAL SOURCE:

25

(A) ORIGIN: Hepatitis C virus

(B) CLONE: J1-1325

30

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

35

G ATC CCA CAA GCT GTC ATG GAC ATG GTG GCG GGG GCC CAC TGG GGA GTC 49

Ile Pro Gln Ala Val Met Asp Met Val Ala Gly Ala His Trp Gly Val

1 5 10 15

40

CTA GCG GGC CTT GCC TAC TAT TCC ATG GTG GGG AAC TGG GCT AAG GTT 97

Leu Ala Gly Leu Ala Tyr Tyr Ser Met Val Gly Asn Trp Ala Lys Val

20 25 30

45

TTG ATT GTG ATG CTA CTC TTT GCC GGC GTT GAC GGG CAT ACC CGC GTG 145

Leu Ile Val Met Leu Leu Phe Ala Gly Val Asp Gly His Thr Arg Val

35 40 45

50

ACG GGG GGG GTG CAA GGC CAT GTC ACC TCT ACA CTC ACG TCC CTC TTT 193

Thr Gly Gly Val Gln Gly His Val Thr Ser Thr Leu Thr Ser Leu Phe

55

50 55 60

5 AGA CCT GGG GCG TCC CAG AAA ATT CAG CTT GTA AAC ACC AAT GGC AGT 241
 Arg Pro Gly Ala Ser Gln Lys Ile Gln Leu Val Asn Thr Asn Gly Ser
 65 70 75 80
 10 TGG CAT ATC AAC AGG ACT GCC CTG AAC TGC AAT GAC TCC CTC AAA ACT 289
 Trp His Ile Asn Arg Thr Ala Leu Asn Cys Asn Asp Ser Leu Lys Thr
 85 90 95
 15 GGG TTT CTT GCC GCG CTG TTC TAC ACA CAC AAG TTC AAC GCG TCC GGA 337
 Gly Phe Leu Ala Ala Leu Phe Tyr Thr His Lys Phe Asn Ala Ser Gly
 100 105 110
 20 TGC CCG GAG CGC ATG GCC AGC TGT CGC TCC ATT GAC AAG TTC GAC CAG 385
 Cys Pro Glu Arg Met Ala Ser Cys Arg Ser Ile Asp Lys Phe Asp Gln
 115 120 125
 25 GGA TGG GGT CCC ATC ACC TAT GCT CAA CCT GAC AAC TCG GAC CAG AGG 433
 Gly Trp Gly Pro Ile Thr Tyr Ala Gln Pro Asp Asn Ser Asp Gln Arg
 130 135 140
 30 CCG TAT TGC TGG CAC TAC GCA CCT CGA CAG TGT GGT ATC GTA CCC GCG 481
 Pro Tyr Cys Trp His Tyr Ala Pro Arg Gln Cys Gly Ile Val Pro Ala
 145 150 155 160
 35 TCG CAG GTG TGC GGT CCA GTG TAT TGC TTC ACC CCA AGC CCT GTT GTA 529
 Ser Gln Val Cys Gly Pro Val Tyr Cys Phe Thr Pro Ser Pro Val Val
 165 170 175
 40 GTG GGG ACG ACC GAT CGT TTC GGC GCC CCT ACG TAT AAC TGG GGG GAC 577
 Val Gly Thr Thr Asp Arg Phe Gly Ala Pro Thr Tyr Asn Trp Gly Asp
 180 185 190
 45 AAT GAG ACG GAC GTG CTG CTC CTA AAC AAC ACG CGG CCG CCG CAT GGC 625
 Asn Glu Thr Asp Val Leu Leu Leu Asn Asn Thr Arg Pro Pro His Gly
 195 200 205
 50 AAC TGG TTC GGC TGT ACA TGG ATG AAT AGC ACT GGG TTC ACC AAG ACG 673
 Asn Trp Phe Gly Cys Thr Trp Met Asn Ser Thr Gly Phe Thr Lys Thr
 55

	210	215	220	
	TGC GGA GGC CCC CCG	TGT AAC ATC AGG GGG	GTC GGC AAC AAC ACC TTG	721
5	Cys Gly Gly Pro Pro	Cys Asn Ile Arg Gly Val	Gly Asn Asn Thr Leu	
	225	230	235	240
10	ACC TGC CCC ACG GAC	TGC TTC CGG AAG CAC	CCC GAC GCC ACT TAC ACA	769
	Thr Cys Pro Thr Asp	Cys Phe Arg Lys His	Pro Asp Ala Thr Tyr Thr	
	245	250	255	
15	AAA TGT GGT TCG GGC	CCT TGG TTG ACA CCT	AGG TGC TTG GTT GAC TAC	817
	Lys Cys Gly Ser Gly	Pro Trp Leu Thr Pro	Arg Cys Leu Val Asp Tyr	
	260	265	270	
20	CCA TAC AGG CTC TGG	CAC TAC CCC TGC ACT	GTC AAC TTT ACC ATC TTC	865
	Pro Tyr Arg Leu Trp	His Tyr Pro Cys Thr	Val Asn Phe Thr Ile Phe	
	275	280	285	
25	AAG GTT AGG ATG TAT	GTG GGG GGC GTG GAG	CAC AGG CTT GAT GCT GCA	913
	Lys Val Arg Met Tyr	Val Gly Gly Val Glu	His Arg Leu Asp Ala Ala	
	290	295	300	
30	TGC AAC TGG ACT CGA	GGA GAG CGT TGC GAC	TTG GAG GAC AGG GAT AGA	961
	Cys Asn Trp Thr Arg	Gly Glu Arg Cys Asp	Leu Glu Asp Arg Asp Arg	
	305	310	315	320
35	GCA GAG CTC AGC CCG	CTA CTG CTG TCT ACG	ACA GAG TGG CAG GTA CTG	1009
	Ala Glu Leu Ser Pro	Leu Leu Leu Ser Thr	Thr Glu Trp Gln Val Leu	
	325	330	335	
40	CCC TGT TCC TTC ACC	ACC CTA CCG GCT CTG	TCC ACT GGT CTA ATC CAT	1057
	Pro Cys Ser Phe Thr	Thr Leu Pro Ala Leu	Ser Thr Gly Leu Ile His	
	340	345	350	
45	CTC CAT CAG AAC GTC	GTG GAC GTG CAA TAC	CTG TAC GGT ATA GGG TCA	1105
	Leu His Gln Asn Val	Val Asp Val Gln Tyr	Leu Tyr Gly Ile Gly Ser	
50	355	360	365	
	GCA GTT GTC TCC TTT	GTA ATC AAA TGG GAG	TAT GTC CTG TTG CTT TTC	1153

55

	Ala Val Val Ser Phe Val Ile Lys Trp Glu Tyr Val Leu Leu Leu Phe	
	370 375 380	
5	CTT CTC CTG GCT GAC GCA CGC GTC TGT GCC TGC TTG TGG ATG ATG CTG	1201
	Leu Leu Leu Ala Asp Ala Arg Val Cys Ala Cys Leu Trp Met Met Leu	
10	385 390 395 400	
	CTG ATA	1207
	Leu Ile	
15		
20		
25		
30		
35		
40		
45		
50		
55		

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1207 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(iv) ANTI-SENSE: No

(vi) ORIGINAL SOURCE:

(A) ORIGIN: Hepatitis C virus

(B) CLONE: N27MX24A-2

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

30	G ATC CCA CAA GCC GTG GTG GAT ATG GTG GCA GGG GCC CAC TGG GGA GTC	49
	Ile Pro Gln Ala Val Val Asp Met Val Ala Gly Ala His Trp Gly Val	
	1 5 10 15	
35	CTG GCG GGC CTT GCC TAC TAT TCC ATG GTG GGG AAC TGG GCT AAG GTC	97
	Leu Ala Gly Leu Ala Tyr Tyr Ser Met Val Gly Asn Trp Ala Lys Val	
	20 25 30	
40	TTG GTT GTG ATG CTG CTC TTC GCC GGT GTT GAC GGG GGG ACC CAC GTG	145
	Leu Val Val Met Leu Leu Phe Ala Gly Val Asp Gly Gly Thr His Val	
	35 40 45	
45	ACA GGG GGG AAG GTA GCC TAC ACC ACC CAG GGC TTT ACA CCC TTC TTT	193
	Thr Gly Gly Lys Val Ala Tyr Thr Thr Gln Gly Phe Thr Pro Phe Phe	
	50 55 60	
50	TCA CGA GGG CCG TCT CAG AAA ATC CAA CTT GTA AAC ACT AAC GGC AGC	241

22

15

	TGC GGG GGC CCC CCG TGC AAC ATC GGG GGG GTC GGC AAC AAT ACC TTG	721
5	Cys Gly Gly Pro Pro Cys Asn Ile Gly Gly Val Gly Asn Asn Thr Leu	
	225 230 235 240	
	ACT TGC CCC ACG GAC TGC TTC CGG AAG CAC CCC GAG GCC ACT TAC ACA	769
10	Thr Cys Pro Thr Asp Cys Phe Arg Lys His Pro Glu Ala Thr Tyr Thr	
	245 250 255	
	AAA TGT GGT TCG GGG CCT TGG TTG ACG CCT AGG TGC CTA GTT CAT TAC	817
15	Lys Cys Gly Ser Gly Pro Trp Leu Thr Pro Arg Cys Leu Val His Tyr	
	260 265 270	
	CCA TAC AGG CTC TGG CAC TAT CCC TGC ACT GTC AAC TTT ACC ATC TTC	865
20	Pro Tyr Arg Leu Trp His Tyr Pro Cys Thr Val Asn Phe Thr Ile Phe	
	275 280 285	
	AAG GTT AGG ATG TAT GTG GGG GGC GTG GAA CAC AGG CTT GAA GCT GCA	913
25	Lys Val Arg Met Tyr Val Gly Gly Val Glu His Arg Leu Glu Ala Ala	
	290 295 300	
	TGC AAT TGG ACC CGA GGA GAG CGT TGT GAC TTG GAG GAC AGG GAT AGA	961
30	Cys Asn Trp Thr Arg Gly Glu Arg Cys Asp Leu Glu Asp Arg Asp Arg	
	305 310 315 320	
	TCA GAG CTT AGC CCG CTA TTG CTG TCC ACA ACA GAG TGG CAG GTA CTG	1009
35	Ser Glu Leu Ser Pro Leu Leu Leu Ser Thr Thr Glu Trp Gln Val Leu	
	325 330 335	
	CCC TGT TCC TTC ACC ACC CTG CCG GCT CTG TCC ACT GGT TTG ATT CAT	1057
40	Pro Cys Ser Phe Thr Thr Leu Pro Ala Leu Ser Thr Gly Leu Ile His	
	340 345 350	
	CTC CAT CAG AAC ATC GTG GAC GTG CAA TAT CTG TAC GGC ATA GGG TCG	1105
45	Leu His Gln Asn Ile Val Asp Val Gln Tyr Leu Tyr Gly Ile Gly Ser	
	355 360 365	
	GCG GTT GTC TCC TTC GCA ATC AAA TGG GAA TAT ATT CTG TTG CTT TTC	1153
50	Ala Val Val Ser Phe Ala Ile Lys Trp Glu Tyr Ile Leu Leu Leu Phe	
55		

370 375 380
 CTC CTC CTG GCG GAC GCG CGC GTC TGT GCC TGC TTG TGG ATG ATG CTG 1201
 5 Leu Leu Leu Ala Asp Ala Arg Val Cys Ala Cys Leu Trp Met Met Leu
 385 390 395 400
 CTG ATA 1207
 10 Leu Ile

(2) INFORMATION FOR SEQ ID NO:3:
 15

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 402 amino acids
 20
 (B) TYPE: protein

(vi) ORIGINAL SOURCE:

(A) ORIGIN: Hepatitis C virus
 25
 (B) CLONE: N27,N19,H19,Y19,MX24

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

35 Ile Pro Gln Ala Val Val Asp Met Val Ala Gly Ala His Trp Gly Val
 1 5 10 15
 40 Leu Ala Gly Leu Ala Tyr Tyr Ser Met Val Gly Asn Trp Ala Lys Val
 20 25 30
 Leu Val Val Met Leu Leu Phe Ala Gly Val Asp Gly Gly Thr His Val
 45 Arg
 35 40 45
 50 Thr Gly Gly Lys Val Ala Tyr Thr Thr Gln Gly Phe Thr Pro Phe Phe
 Arg Ser

55

Ser

50 55 60

5 Ser Arg Gly Pro Ser Gln Lys Ile Gln Leu Val Asn Thr Asn Gly Ser

Arg

65 70 75 80

10 Trp His Ile Asn Arg Thr Ala Leu Asn Cys Asn Asp Ser Leu Asn Thr

Gln

85 90 95

15 Gly Phe Leu Ala Ala Leu Phe Tyr Thr His Ser Phe Asn Ala Ser Gly

Thr Arg Asp

100 105 110

20 Cys Pro Glu Arg Met Ala Gly Cys Arg Pro Ile Asp Glu Phe Ala Gln

Ser Ser

115 120 125

25 Gly Trp Gly Pro Ile Thr His Val Val Pro Asn Ile Ser Asp Gln Arg

Asp Asp Val

130 135 140

30 Pro Tyr Cys Trp His Tyr Ala Pro Arg Pro Cys Gly Ile Val Pro Ala

Val

145 150 155 160

35 Ser Gln Val Cys Gly Pro Val Tyr Cys Phe Thr Pro Ser Pro Val Val

Trp

165 170 175

40 Val Gly Thr Thr Asp Arg Phe Gly Ala Pro Thr Tyr Asn Trp Gly Asn

Ser Thr Ala

180 185 190

45 Asn Glu Thr Asp Val Leu Leu Leu Asn Asn Thr Arg Pro Pro Gln Gly

195 200 205

50 Asn Trp Phe Gly Cys Thr Trp Met Asn Gly Thr Gly Phe Thr Lys Thr

55

210 215 220
 Cys Gly Gly Pro Pro Cys Asn Ile Gly Gly Val Gly Asn Asn Thr Leu
 5 225 230 235 240
 Thr Cys Pro Thr Asp Cys Phe Arg Lys His Pro Glu Ala Thr Tyr Thr
 245 250 255
 10 Lys Cys Gly Ser Gly Pro Trp Leu Thr Pro Arg Cys Leu Val His Tyr
 260 265 270
 Pro Tyr Arg Leu Trp His Tyr Pro Cys Thr Val Asn Phe Thr Ile Phe
 15 275 280 285
 Lys Val Arg Met Tyr Val Gly Gly Val Glu His Arg Leu Glu Ala Ala
 290 295 300
 20 Cys Asn Trp Thr Arg Gly Glu Arg Cys Asp Leu Glu Asp Arg Asp Arg
 305 310 315 320
 25 Ser Glu Leu Ser Pro Leu Leu Leu Ser Thr Thr Glu Trp Gln Val Leu
 325 330 335
 Pro Cys Ser Phe Thr Thr Leu Pro Ala Leu Ser Thr Gly Leu Ile His
 340 345 350
 30 Leu His Gln Asn Ile Val Asp Val Gln Tyr Leu Tyr Gly Ile Gly Ser
 355 360 365
 35 Ala Val Val Ser Phe Ala Ile Lys Trp Glu Tyr Ile Leu Leu Leu Phe
 370 375 380
 Leu Leu Leu Ala Asp Ala Arg Val Cys Ala Cys Leu Trp Met Met Leu
 40 385 390 395 400
 Leu Ile

45 (2) INFORMATION FOR SEQ ID NO:4:

50 (i) SEQUENCE CHARACTERISTICS:

 (A) LENGTH: 1207 base pairs

55

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(iv) ANTI-SENSE: No

(vi) ORIGINAL SOURCE:

(A) ORIGIN: Hepatitis C virus

(B) CLONE: BK164

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

G	ATC	CCA	CAA	GCC	GTC	GTG	GAC	ATG	GTG	GCG	GGG	GCC	CAC	TGG	GGA	GTC	49
Ile	Pro	Gln	Ala	Val	Val	Asp	Met	Val	Ala	Gly	Ala	His	Trp	Gly	Val		
1				5					10					15			
CTG	GCG	GGC	CTT	GCC	TAC	TAT	TCC	ATG	GCG	GGG	AAC	TGG	GCT	AAG	GTT	97	
Leu	Ala	Gly	Leu	Ala	Tyr	Tyr	Ser	Met	Ala	Gly	Asn	Trp	Ala	Lys	Val		
			20				25						30				
CTG	ATT	GTG	ATG	CTA	CTT	TTT	GCT	GGC	GTT	GAC	GGG	GAT	ACC	CAC	GTG	145	
Leu	Ile	Val	Met	Leu	Leu	Phe	Ala	Gly	Val	Asp	Gly	Asp	Thr	His	Val		
		35				40						45					
ACA	GGG	GGG	GCG	CAA	GCC	AAA	ACC	ACC	AAC	AGG	CTC	GTG	TCC	ATG	TTC	193	
Thr	Gly	Gly	Ala	Gln	Ala	Lys	Thr	Thr	Asn	Arg	Leu	Val	Ser	Met	Phe		
		50				55						60					
GCA	AGT	GGG	CCG	TCT	CAG	AAA	ATC	CAG	CTT	ATA	AAC	ACC	AAT	GGG	AGT	241	
Ala	Ser	Gly	Pro	Ser	Gln	Lys	Ile	Gln	Leu	Ile	Asn	Thr	Asn	Gly	Ser		
		65				70				75				80			
TGG	CAC	ATC	AAC	AGG	ACT	GCC	CTG	AAC	TGC	AAT	GAC	TCT	CTC	CAG	ACT	289	
Trp	His	Ile	Asn	Arg	Thr	Ala	Leu	Asn	Cys	Asn	Asp	Ser	Leu	Gln	Thr		

	85	90	95	
	GGG TTT CTT GCC GCG CTG TTC TAC ACA CAT AGT TTC AAC TCG TCC GGG	337		
5	Gly Phe Leu Ala Ala Leu Phe Tyr Thr His Ser Phe Asn Ser Ser Gly			
	100	105	110	
	TGC CCA GAG CGC ATG GCC CAG TGC CGC ACC ATT GAC AAG TTC GAC CAG	385		
10	Cys Pro Glu Arg Met Ala Gln Cys Arg Thr Ile Asp Lys Phe Asp Gln			
	115	120	125	
	GGA TGG GGT CCC ATT ACT TAT GCT GAG TCT AGC AGA TCA GAC CAG AGG	433		
15	Gly Trp Gly Pro Ile Thr Tyr Ala Glu Ser Ser Arg Ser Asp Gln Arg			
	130	135	140	
	CCA TAT TGC TGG CAC TAC CCA CCT CCA CAA TGT ACC ATC GTA CCT GCG	481		
20	Pro Tyr Cys Trp His Tyr Pro Pro Pro Gln Cys Thr Ile Val Pro Ala			
	145	150	155	160
	TCG GAG GTG TGC GGC CCA GTG TAC TGC TTC ACC CCA AGC CCT GTC GTC	529		
25	Ser Glu Val Cys Gly Pro Val Tyr Cys Phe Thr Pro Ser Pro Val Val			
	165	170	175	
	GTG GGG ACG ACC GAT CGT TTC GGT GTC CCT ACG TAT AGA TGG GGG GAG	577		
30	Val Gly Thr Thr Asp Arg Phe Gly Val Pro Thr Tyr Arg Trp Gly Glu			
	180	185	190	
	AAC GAG ACT GAC GTG CTG CTG CTC AAC AAC ACG CGG CCG CCA GGC	625		
35	Asn Glu Thr Asp Val Leu Leu Leu Asn Asn Thr Arg Pro Pro Gln Gly			
	195	200	205	
	AAC TGG TTC GGC TGC ACA TGG ATG AAT AGC ACC GGG TTC ACC AAG ACA	673		
40	Asn Trp Phe Gly Cys Thr Trp Met Asn Ser Thr Gly Phe Thr Lys Thr			
	210	215	220	
	TGT GGG GGG CCC CCC TGT AAC ATC GGG GGG GTC GGC AAC AAC ACC CTG	721		
45	Cys Gly Gly Pro Pro Cys Asn Ile Gly Gly Val Gly Asn Asn Thr Leu			
	225	230	235	240
50	ACC TGC CCC ACG GAC TGC TTC CGG AAG CAC CCC GAG GCT ACC TAC ACA	769		
55				

	Thr Cys Pro Thr Asp Cys Phe Arg Lys His Pro Glu Ala Thr Tyr Thr	
	245 250 255	
5	AAA TGT GGT TCG GGG CCT TGG CTG ACA CCT AGG TGC ATG GTT GAC TAT	817
	Lys Cys Gly Ser Gly Pro Trp Leu Thr Pro Arg Cys Met Val Asp Tyr	
	260 265 270	
10	CCA TAC AGG CTC TGG CAT TAC CCC TGC ACT GTT AAC TTT ACC ATC TTC	865
	Pro Tyr Arg Leu Trp His Tyr Pro Cys Thr Val Asn Phe Thr Ile Phe	
	275 280 285	
15	AAG GTT AGG ATG TAT GTG GGG GGG GTG GAG CAC AGG CTC AAT GCT GCA	913
	Lys Val Arg Met Tyr Val Gly Gly Val Glu His Arg Leu Asn Ala Ala	
	290 295 300	
20	TGC AAT TGG ACC CGA GGA GAG CGT TGT GAC TTG GAG GAC AGG GAT AGG	961
	Cys Asn Trp Thr Arg Gly Glu Arg Cys Asp Leu Glu Asp Arg Asp Arg	
	305 310 315 320	
25	CCG GAG CTC AGC CCG CTG CTG CTG TCT ACA ACA GAG TGG CAG GTA CTG	1009
	Pro Glu Leu Ser Pro Leu Leu Leu Ser Thr Thr Glu Trp Gln Val Leu	
	325 330 335	
30	CCC TGT TCC TTC ACC ACC CTA CCA GCT CTG TCC ACT GGC TTG ATT CAC	1057
	Pro Cys Ser Phe Thr Thr Leu Pro Ala Leu Ser Thr Gly Leu Ile His	
	340 345 350	
35	CTC CAT CAG AAC ATC GTG GAC GTG CAA TAC CTA TAC GGT ATA GGG TCA	1105
	Leu His Gln Asn Ile Val Asp Val Gln Tyr Leu Tyr Gly Ile Gly Ser	
	355 360 365	
40	GCG GTT GTC TCC TTT GCA ATC AAA TGG GAG TAT GTC CTG TTG CTT TTC	1153
	Ala Val Val Ser Phe Ala Ile Lys Trp Glu Tyr Val Leu Leu Leu Phe	
	370 375 380	
45	CTT CTC CTA GCG GAC GCA CGT GTC TGT GCC TGC TTG TGG ATG ATG CTG	1201
	Leu Leu Leu Ala Asp Ala Arg Val Cys Ala Cys Leu Trp Met Met Leu	
	385 390 395 400	
50		
55		

CTG ATA

1207

Leu Ile

5

(2) INFORMATION FOR SEQ ID NO:5:

10

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1207 base pairs

15

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

20

(iv) ANTI-SENSE: No

25

(vi) ORIGINAL SOURCE:

(A) ORIGIN: Hepatitis C virus

(B) CLONE: HCV-J

30

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

35

G ATC CCA CAA GCC GTC GTG GAC ATG GTG GCG GGG GCC CAC TGG GGT GTC 49

Ile Pro Gln Ala Val Val Asp Met Val Ala Gly Ala His Trp Gly Val

40

1 5 10 15

CTA GCG GGC CTT GCC TAC TAT TCC ATG GTG GGG AAC TGG GCT AAG GTC 97

Leu Ala Gly Leu Ala Tyr Tyr Ser Met Val Gly Asn Trp Ala Lys Val

45

20 25 30

TTG ATT GTG ATG CTA CTC TTT GCT GGC GTT GAC GGG CAC ACC CAC GTG 145

Leu Ile Val Met Leu Leu Phe Ala Gly Val Asp Gly His Thr His Val

50

35 40 45

ACA GGG GGA AGG GTA GCC TCC AGC ACC CAG AGC CTC GTG TCC TGG CTC 193

55

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24

AAC TGG TTT GGG TGC ACG TGG ATG AAC AGC ACT GGG TTC ACC AAG ACG 673
 Asn Trp Phe Gly Cys Thr Trp Met Asn Ser Thr Gly Phe Thr Lys Thr
 5 210 215 220
 TGC GGG GGC CCT CCG TGC AAC ATC GGG GGG GTC GGC AAC AAC ACC TTG 721
 Cys Gly Gly Pro Pro Cys Asn Ile Gly Gly Val Gly Asn Asn Thr Leu
 10 225 230 235 240
 GTC TGC CCC ACG GAT TGC TTC CGG AAG CAC CCC GAG GCC ACT TAC ACA 769
 Val Cys Pro Thr Asp Cys Phe Arg Lys His Pro Glu Ala Thr Tyr Thr
 15 245 250 255
 AAG TGT GGC TCG GGG CCC TGG TTG ACA CCC AGG TGC ATG GTT GAC TAC 817
 Lys Cys Gly Ser Gly Pro Trp Leu Thr Pro Arg Cys Met Val Asp Tyr
 20 260 265 270
 CCA TAC AGG CTC TGG CAC TAC CCC TGC ACT GTT AAC TTT ACC GTC TTT 865
 Pro Tyr Arg Leu Trp His Tyr Pro Cys Thr Val Asn Phe Thr Val Phe
 25 275 280 285
 AAG GTC AGG ATG TAT GTG GGG GGC GTG GAG CAC AGG CTC AAT GCT GCA 913
 Lys Val Arg Met Tyr Val Gly Gly Val Glu His Arg Leu Asn Ala Ala
 30 290 295 300
 TGC AAT TGG ACT CGA GGA GAG CGC TGT GAC TTG GAG GAC AGG GAT AGG 961
 Cys Asn Trp Thr Arg Gly Glu Arg Cys Asp Leu Glu Asp Arg Asp Arg
 35 305 310 315 320
 TCA GAA CTC AGC CCG CTG CTG CTG TCT ACA ACA GAG TGG CAG ATA CTG 1009
 Ser Glu Leu Ser Pro Leu Leu Leu Ser Thr Thr Glu Trp Gln Ile Leu
 40 325 330 335
 CCC TGT TCC TTC ACC ACC CTA CCG GCC CTG TCC ACT GGC TTG ATC CAT 1057
 Pro Cys Ser Phe Thr Thr Leu Pro Ala Leu Ser Thr Gly Leu Ile His
 45 340 345 350
 CTT CAC CGG AAC ATC GTG GAC GTG CAA TAC CTG TAC GGT ATA GGG TCG 1105
 Leu His Arg Asn Ile Val Asp Val Gln Tyr Leu Tyr Gly Ile Gly Ser
 50
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20 (2) INFORMATION FOR SEQ ID NO:6:

25 (A) LENGTH: 1207 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
30 (D) TOPOLOGY: linear

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40 (B) CLONE: HCV-RNA33

45

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	CTG GCG GGC CTG GCC TAC TAT TCC ATG GTG GGG AAC TGG GCT AAG GTT	97
5	Leu Ala Gly Leu Ala Tyr Tyr Ser Met Val Gly Asn Trp Ala Lys Val	
	20 25 30	
	TTG ATT GTG ATG CTA CTC TTT GCC GGC GTT GAC GGG CAA ACC TAT ACG	145
10	Leu Ile Val Met Leu Leu Phe Ala Gly Val Asp Gly Gln Thr Tyr Thr	
	35 40 45	
	ACG GGG GGC GCG GTT GCC CGC ACC ACC ACC GGG TTC GCG TCC CTC TTC	193
15	Thr Gly Gly Ala Val Ala Arg Thr Thr Thr Gly Phe Ala Ser Leu Phe	
	50 55 60	
	TCC GCT GGG TCG CAG GAG AAC ATC CAG CTT ATA AAC ACC AAT GGC AGC	241
20	Ser Ala Gly Ser Gln Glu Asn Ile Gln Leu Ile Asn Thr Asn Gly Ser	
	65 70 75 80	
	TGG CAC ATC AAC AGG ACT GCC CTG AAC TGC AAC GAC TCC CTC AAC ACT	289
25	Trp His Ile Asn Arg Thr Ala Leu Asn Cys Asn Asp Ser Leu Asn Thr	
	85 90 95	
	GGA TTT CTT GCC GCG CTG TTC TAC ACA CAC AAG TTC AAC TCA TCC AGA	337
30	Gly Phe Leu Ala Ala Leu Phe Tyr Thr His Lys Phe Asn Ser Ser Arg	
	100 105 110	
	GCC GAG AGC GTA TTG GCC AGC TGC CGC TTC ATC GAC GAG TTC GAT CAG	385
35	Ala Glu Ser Val Leu Ala Ser Cys Arg Phe Ile Asp Glu Phe Asp Gln	
	115 120 125	
	GGA TGG GGC CCC ATC ACT TAC ACC GAG CGT AAC AGT TCG GAC CAG AGG	433
40	Gly Trp Gly Pro Ile Thr Tyr Thr Glu Arg Asn Ser Ser Asp Gln Arg	
	130 135 140	
	CCT TAT TGC TGG CAC TAT CCA CCC CGA CAG TGT GGT ATC ATA CCC GCG	481
45	Pro Tyr Cys Trp His Tyr Pro Pro Arg Gln Cys Gly Ile Ile Pro Ala	
	145 150 155 160	
	TCG GAG GTG TGC GGT CCA GTG TAT TGT TTC ACC CCA AGC CCT GTT GTG	529
50	Ser Glu Val Cys Gly Pro Val Tyr Cys Phe Thr Pro Ser Pro Val Val	

55

	165	170	175	
	GTG GGG ACA ACC GAT CGG TTC GGT GTC CCT ACA TAC AGC TGG GGG GAG	577		
5	Val Gly Thr Thr Asp Arg Phe Gly Val Pro Thr Tyr Ser Trp Gly Glu			
	180	185	190	
10	AAT GAG ACG GAC GTG CTG GTT CTC AAC AAC ACG CGG CCG CCG CAG GGC	625		
	Asn Glu Thr Asp Val Leu Val Leu Asn Asn Thr Arg Pro Pro Gln Gly			
	195	200	205	
15	AAC TGG TTC GGC TGT ACA TGG ATG AAT GGC ACT GGT TTC ACC AAG ACA	673		
	Asn Trp Phe Gly Cys Thr Trp Met Asn Gly Thr Gly Phe Thr Lys Thr			
	210	215	220	
20	TGC GGG GGT CCC CCG TGT CAC ATC GGG GGG CGC GGC AAC AAC ACC CTG	721		
	Cys Gly Gly Pro Pro Cys His Ile Gly Gly Arg Gly Asn Asn Thr Leu			
	225	230	235	240
25	ACT TGC CCC ACG GAC TGC TTC CGG AAG CAT CCC GAG GCT ACG TAT ACA	769		
	Thr Cys Pro Thr Asp Cys Phe Arg Lys His Pro Glu Ala Thr Tyr Thr			
	245	250	255	
30	AAA TGT GGT TCG GGG CCT TGG TTG ACA CCT AGG TGC ATG GTT GAT TAC	817		
	Lys Cys Gly Ser Gly Pro Trp Leu Thr Pro Arg Cys Met Val Asp Tyr			
	260	265	270	
35	CCA TAC AGG CTC TGG CAC TAC CCC TGC ACT GTC AAC TTT ACC ACC TTT	865		
	Pro Tyr Arg Leu Trp His Tyr Pro Cys Thr Val Asn Phe Thr Thr Phe			
	275	280	285	
40	AAG GTT AGG ATG TAT GTG GGG GGC GTG GAG CAC AGG CTC ATT GCT GCA	913		
	Lys Val Arg Met Tyr Val Gly Gly Val Glu His Arg Leu Ile Ala Ala			
	290	295	300	
45	TGC AAT TGG ACT CGA GGA GAC CGT TGT AAC TTG GAG GAC AGG GAT AGA	961		
	Cys Asn Trp Thr Arg Gly Asp Arg Cys Asn Leu Glu Asp Arg Asp Arg			
50	305	310	315	320
	TCA GAG CTT AGT CCG CTG CTG CTG TCT ACG ACA GAG TGG CAG ATA CTG	1009		

55

Ser Glu Leu Ser Pro Leu Leu Leu Ser Thr Thr Glu Trp Gln Ile Leu
 325 330 335
 5 CCC TGT TCC TTC ACC ACC CTA CCG GCT CTC TCC ACC GGT TTG ATC CAT 1057
 Pro Cys Ser Phe Thr Thr Leu Pro Ala Leu Ser Thr Gly Leu Ile His
 340 345 350
 10 CTC CAT CAG AAC ATC GTG GAC GTG CAA TAC CTG TAC GGT ATA GGG TCT 1105
 Leu His Gln Asn Ile Val Asp Val Gln Tyr Leu Tyr Gly Ile Gly Ser
 355 360 365
 15 GCT GTT GTC TCC ATT GCA ATC AGG TGG GAA TAT GTC CTG TTG CTT TTC 1153
 Ala Val Val Ser Ile Ala Ile Arg Trp Glu Tyr Val Leu Leu Leu Phe
 370 375 380
 20 CTT CTC CTG GCG GAC GCG CGT GTC TGT GCC TGC TTG TGG ATG ATG CTG 1201
 Leu Leu Leu Ala Asp Ala Arg Val Cys Ala Cys Leu Trp Met Met Leu
 385 390 395 400
 25 CTG ATA 1207
 Leu Ile

30

(2) INFORMATION FOR SEQ ID NO:7:

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(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1207 base pairs

(B) TYPE: nucleic acid

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(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

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(iv) ANTI-SENSE: No

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(vi) ORIGINAL SOURCE:

(A) ORIGIN: Hepatitis C virus

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(B) CLONE: HCV1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

5 G ATC CCA CAA GCC ATC TTG GAC ATG ATC GCT GGT GCT CAC TGG GGA GTC 49
 10 Ile Pro Gln Ala Ile Leu Asp Met Ile Ala Gly Ala His Trp Gly Val
 1 5 10 15
 15 CTG GCG GGC ATA GCG TAT TTC TCC ATG GTG GGG AAC TGG GCG AAG GTC 97
 Leu Ala Gly Ile Ala Tyr Phe Ser Met Val Gly Asn Trp Ala Lys Val
 20 25 30
 20 CTG GTA GTG CTG CTG CTA TTT GCC GGC GTC GAC GCG GAA ACC CAC GTC 145
 Leu Val Val Leu Leu Leu Phe Ala Gly Val Asp Ala Glu Thr His Val
 35 40 45
 25 ACC GGG GGA AGT GCC GGC CAC ACT GTG TCT GGA TTT GTT AGC CTC CTC 193
 Thr Gly Gly Ser Ala Gly His Thr Val Ser Gly Phe Val Ser Leu Leu
 50 55 60
 30 GCA CCA GGC GCC AAG CAG AAC GTC CAG CTG ATC AAC ACC AAC GGC AGT 241
 Ala Pro Gly Ala Lys Gln Asn Val Gln Leu Ile Asn Thr Asn Gly Ser
 65 70 75 80
 35 TGG CAC CTC AAT AGC ACG GCC CTG AAC TGC AAT GAT AGC CTC AAC ACC 289
 Trp His Leu Asn Ser Thr Ala Leu Asn Cys Asn Asp Ser Leu Asn Thr
 85 90 95
 40 GGC TGG TTG GCA GGG CTT TTC TAT CAC CAC AAG TTC AAC TCT TCA GGC 337
 Gly Trp Leu Ala Gly Leu Phe Tyr His His Lys Phe Asn Ser Ser Gly
 100 105 110
 45 TGT CCT GAG AGG CTA GCC AGC TGC CGA CCC CTT ACC GAT TTT GAC CAG 385
 Cys Pro Glu Arg Leu Ala Ser Cys Arg Pro Leu Thr Asp Phe Asp Gln
 115 120 125
 50 GGC TGG GGC CCT ATC AGT TAT GCC AAC GGA AGC GGC CCC GAC CAG CGC 433

55

	Gly Trp Gly Pro Ile Ser Tyr Ala Asn Gly Ser Gly Pro Asp Gln Arg	
	130 135 140	
5	CCC TAC TGC TGG CAC TAC CCC CCA AAA CCT TGC GGT ATT GTG CCC GCG	481
	Pro Tyr Cys Trp His Tyr Pro Pro Lys Pro Cys Gly Ile Val Pro Ala	
	145 150 155 160	
10	AAG AGT GTG TGT GGT CCG GTA TAT TGC TTC ACT CCC AGC CCC GTG GTG	529
	Lys Ser Val Cys Gly Pro Val Tyr Cys Phe Thr Pro Ser Pro Val Val	
	165 170 175	
15	GTG GGA ACG ACC GAC AGG TCG GGC GCG CCC ACC TAC AGC TGG GGT GAA	577
	Val Gly Thr Thr Asp Arg Ser Gly Ala Pro Thr Tyr Ser Trp Gly Glu	
	180 185 190	
20	AAT GAT ACG GAC GTC TTC GTC CTT AAC AAT ACC AGG CCA CCG CTG GGC	625
	Asn Asp Thr Asp Val Phe Val Leu Asn Asn Thr Arg Pro Pro Leu Gly	
	195 200 205	
25	AAT TGG TTC GGT TGT ACC TGG ATG AAC TCA ACT GGA TTC ACC AAA GTG	673
	Asn Trp Phe Gly Cys Thr Trp Met Asn Ser Thr Gly Phe Thr Lys Val	
	210 215 220	
30	TGC GGA GCG CCT CCT TGT GTC ATC GGA GGG GCG GGC AAC AAC ACC CTG	721
	Cys Gly Ala Pro Pro Cys Val Ile Gly Gly Ala Gly Asn Asn Thr Leu	
	225 230 235 240	
35	CAC TGC CCC ACT GAT TGC TTC CGC AAG CAT CCG GAC GCC ACA TAC TCT	769
	His Cys Pro Thr Asp Cys Phe Arg Lys His Pro Asp Ala Thr Tyr Ser	
	245 250 255	
40	CGG TGC GGC TCC GGT CCC TGG ATC ACA CCC AGG TGC CTG GTC GAC TAC	817
	Arg Cys Gly Ser Gly Pro Trp Ile Thr Pro Arg Cys Leu Val Asp Tyr	
	260 265 270	
45	CCG TAT AGG CTT TGG CAT TAT CCT TGT ACC ATC AAC TAC ACC ATA TTT	865
	Pro Tyr Arg Leu Trp His Tyr Pro Cys Thr Ile Asn Tyr Thr Ile Phe	
	275 280 285	
50		
55		

AAA ATC AGG ATG TAC GTG GGA GGG GTC GAA CAC AGG CTG GAA GCT GCC 913
 Lys Ile Arg Met Tyr Val Gly Gly Val Glu His Arg Leu Glu Ala Ala
 5 290 295 300
 TGC AAC TGG ACG CGG GGC GAA CGT TGC GAT CTG GAA GAC AGG GAC AGG 961
 Cys Asn Trp Thr Arg Gly Glu Arg Cys Asp Leu Glu Asp Arg Asp Arg
 10 305 310 315 320
 TCC GAG CTC AGC CCG TTA CTG CTG ACC ACT ACA CAG TGG CAG GTC CTC 1009
 Ser Glu Leu Ser Pro Leu Leu Leu Thr Thr Thr Gln Trp Gln Val Leu
 15 325 330 335
 CCG TGT TCC TTC ACA ACC CTA CCA GCC TTG TCC ACC GGC CTC ATC CAC 1057
 Pro Cys Ser Phe Thr Thr Leu Pro Ala Leu Ser Thr Gly Leu Ile His
 20 340 345 350
 CTC CAC CAG AAC ATT GTG GAC GTG CAG TAC TTG TAC GGG GTG GGG TCA 1105
 Leu His Gln Asn Ile Val Asp Val Gln Tyr Leu Tyr Gly Val Gly Ser
 25 355 360 365
 AGC ATC GCG TCC TGG GCC ATT AAG TGG GAG TAC GTC GTT CTC CTG TTC 1153
 Ser Ile Ala Ser Trp Ala Ile Lys Trp Glu Tyr Val Val Leu Leu Phe
 30 370 375 380
 CTT CTG CTT GCA GAC GCG CGC GTC TGC TCC TGC TTG TGG ATG ATG CTA 1201
 Leu Leu Leu Ala Asp Ala Arg Val Cys Ser Cys Leu Trp Met Met Leu
 35 385 390 395 400
 CTC ATA 1207
 40 Leu Ile
 45
 50
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(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1207 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(iv) ANTI-SENSE: No

(vi) ORIGINAL SOURCE:

(A) ORIGIN: Hepatitis C virus

(B) CLONE: H77

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

G ATC CCA CAA GCC ATC ATG GAC ATG ATC GCT GGT GCT CAC TGG GGA GTC 49
 Ile Pro Gln Ala Ile Met Asp Met Ile Ala Gly Ala His Trp Gly Val
 1 5 10 15
 CTG GCG GGC ATA GCG TAT TTC TCC ATG GTG GGG AAC TGG GCG AAG GTC 97
 Leu Ala Gly Ile Ala Tyr Phe Ser Met Val Gly Asn Trp Ala Lys Val
 20 25 30
 CTG GTA GTG CTG CTG CTA TTT GCC GGC GTC GAC GCG GAA ACC CAC GTC 145
 Leu Val Val Leu Leu Leu Phe Ala Gly Val Asp Ala Glu Thr His Val
 35 40 45
 ACC GGG GGA AGT GCC GGC CGC ACC ACG GCT GGG CTT GTT GGT CTC CTT 193
 Thr Gly Gly Ser Ala Gly Arg Thr Thr Ala Gly Leu Val Gly Leu Leu
 50 55 60
 ACA CCA GGC GCC AAG CAG AAC ATC CAA CTG ATC AAC ACC AAC GGC AGT 241

5 Thr Pro Gly Ala Lys Gln Asn Ile Gln Leu Ile Asn Thr Asn Gly Ser
 65 70 75 80
 TGG CAC ATC AAT AGC ACG GCC TTG AAC TGC AAT GAA AGC CTT AAC ACC 289
 10 Trp His Ile Asn Ser Thr Ala Leu Asn Cys Asn Glu Ser Leu Asn Thr
 85 90 95
 GGC TGG TTA GCA GGG CTC TTC TAT CAC CAC AAA TTC AAC TCT TCA GGC 337
 15 Gly Trp Leu Ala Gly Leu Phe Tyr His His Lys Phe Asn Ser Ser Gly
 100 105 110
 TGT CCT GAG AGG TTG GCC AGC TGC CGA CGC CTT ACC GAT TTT GCC CAG 385
 20 Cys Pro Glu Arg Leu Ala Ser Cys Arg Arg Leu Thr Asp Phe Ala Gln
 115 120 125
 GGC TGG GGT CCT ATC AGT TAT GCC AAC GGA AGC GGC CTC GAC GAA CGC 433
 25 Gly Trp Gly Pro Ile Ser Tyr Ala Asn Gly Ser Gly Leu Asp Glu Arg
 130 135 140
 CCC TAC TGC TGG CAC TAC CCT CCA AGA CCT TGT GGC ATT GTG CCC GCA 481
 30 Pro Tyr Cys Trp His Tyr Pro Pro Arg Pro Cys Gly Ile Val Pro Ala
 145 150 155 160
 AAG AGC GTG TGT GGC CCG GTA TAT TGC TTC ACT CCC AGC CCC GTG GTG 529
 35 Lys Ser Val Cys Gly Pro Val Tyr Cys Phe Thr Pro Ser Pro Val Val
 165 170 175
 GTG GGA ACG ACC GAC AGG TCG GGC GCG CCT ACC TAC AGC TGG GGT GCA 577
 40 Val Gly Thr Thr Asp Arg Ser Gly Ala Pro Thr Tyr Ser Trp Gly Ala
 180 185 190
 AAT GAT ACG GAT GTC TTC GTC CTT AAC AAC ACC AGG CCA CCG CTG GGC 625
 45 Asn Asp Thr Asp Val Phe Val Leu Asn Asn Thr Arg Pro Pro Leu Gly
 195 200 205
 AAT TGG TTC GGT TGT ACC TGG ATG AAC TCA ACT GGA TTC ACC AAA GTG 673
 50 Asn Trp Phe Gly Cys Thr Trp Met Asn Ser Thr Gly Phe Thr Lys Val
 210 215 220

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370 375 380
 CTT CTG CTT GCA GAC GCG CGC GTC TGC TCC TGC TTG TGG ATG ATG TTA 1201
 5 Leu Leu Leu Ala Asp Ala Arg Val Cys Ser Cys Leu Trp Met Met Leu
 385 390 395 400
 CTC ATA 1207
 10 Leu Ile

15 (2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

20 (A) LENGTH: 1207 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 25 (D) TOPOLOGY: linear

(iv) ANTI-SENSE: No

30

(vi) ORIGINAL SOURCE:

35 (A) ORIGIN: Hepatitis C virus
 (B) CLONE: H90

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

40

G ATC CCA CAA GCC ATC ATG GAT ATG ATC GCT GGT GCT CAC TGG GGA GTC 49
 45 Ile Pro Gln Ala Ile Met Asp Met Ile Ala Gly Ala His Trp Gly Val
 1 5 10 15
 CTG GCG GGC ATA GCG TAT TTC TCC ATG GTA GGG AAC TGG GCG AAG GTC 97
 50 Leu Ala Gly Ile Ala Tyr Phe Ser Met Val Gly Asn Trp Ala Lys Val
 20 25 30

55

CTA GTA GTG CTG CTG CTA TTT GCC GGC GTC GAC GCG GAA ACC CAC GTC 145
 Leu Val Val Leu Leu Leu Phe Ala Gly Val Asp Ala Glu Thr His Val
 5 35 40 45
 ACC GGG GGA AGT GCC GGC CGC TCC GTG CTT GGG ATT GCT AGT TTC CTT 193
 Thr Gly Gly Ser Ala Gly Arg Ser Val Leu Gly Ile Ala Ser Phe Leu
 10 50 55 60
 ACA CGA GGC CCC AAG CAG AAC ATC CAG CTG ATC AAA ACC AAC GGC AGT 241
 Thr Arg Gly Pro Lys Gln Asn Ile Gln Leu Ile Lys Thr Asn Gly Ser
 15 65 70 75 80
 TGG CAC ATC AAT AGC ACG GCC CTG AAC TGC AAT GAC AGC CTT AAC GCC 289
 Trp His Ile Asn Ser Thr Ala Leu Asn Cys Asn Asp Ser Leu Asn Ala
 20 85 90 95
 GGC TGG ATA GCG GGG CTC TTC TAT CAC CAT GGA TTC AAC TCT TCA GGC 337
 Gly Trp Ile Ala Gly Leu Phe Tyr His His Gly Phe Asn Ser Ser Gly
 25 100 105 110
 TGT CCT GAG AGG TTG GCC AGC TGC CGA CGC CTT ACC GAT TTT GAC CAG 385
 Cys Pro Glu Arg Leu Ala Ser Cys Arg Arg Leu Thr Asp Phe Asp Gln
 30 115 120 125
 GGC TGG GGC CCT ATC AGT TAT GCC AAC GGA AGC GGC CCC GAC GAA CGT 433
 Gly Trp Gly Pro Ile Ser Tyr Ala Asn Gly Ser Gly Pro Asp Glu Arg
 35 130 135 140
 CCC TAC TGC TGG CAC TAC CCC CCA AGA CCT TGT GGC ATT GTG CCC GCA 481
 Pro Tyr Cys Trp His Tyr Pro Pro Arg Pro Cys Gly Ile Val Pro Ala
 40 145 150 155 160
 AAG AGC GTG TGT GGC CCG GTA TAC TGC TTC ACT CCC AGC CCC GTG GTG 529
 Lys Ser Val Cys Gly Pro Val Tyr Cys Phe Thr Pro Ser Pro Val Val
 45 165 170 175
 GTG GGA ACG ACC GAC AGG TCG GGC GCG CCT ACC TAC AAC TGG GGT GAA 577
 Val Gly Thr Thr Asp Arg Ser Gly Ala Pro Thr Tyr Asn Trp Gly Glu
 50
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	180	185	190	
	AAT GAT ACG GAT GTC CTC ATC CTT AAC AAC ACC AGG CCG CCG CTG GGC	625		
5	Asn Asp Thr Asp Val Leu Ile Leu Asn Asn Thr Arg Pro Pro Leu Gly			
	195	200	205	
	AAT TGG TTC GGT TGT ACC TGG ATG AAC TCA ACT GGA TTC ACC AAA GTG	673		
10	Asn Trp Phe Gly Cys Thr Trp Met Asn Ser Thr Gly Phe Thr Lys Val			
	210	215	220	
	TGC GGA GCG CCC CCT TGT GTC ATC GGA GGG GTG GGC AAC AAC ACC TTG	721		
15	Cys Gly Ala Pro Pro Cys Val Ile Gly Gly Val Gly Asn Asn Thr Leu			
	225	230	235	240
	CGC TGC CCC ACT GAT TGT TTC CGC AAG CAT CCG GAA GCC ACA TAC TCT	769		
20	Arg Cys Pro Thr Asp Cys Phe Arg Lys His Pro Glu Ala Thr Tyr Ser			
	245	250	255	
	CGG TGC GGC TCC GGT CCC TGG ATC ACA CCC AGG TGC ATG GTC CAC TAC	817		
25	Arg Cys Gly Ser Gly Pro Trp Ile Thr Pro Arg Cys Met Val His Tyr			
	260	265	270	
	CCG TAT AGG CTT TGG CAC TAT CCT TGT ACC ATC AAT TAC ACT ATA TTT	865		
30	Pro Tyr Arg Leu Trp His Tyr Pro Cys Thr Ile Asn Tyr Thr Ile Phe			
	275	280	285	
	AAA GTC AGG ATG TAC GTG GGA GGG ATC GAG CAC AGG CTG GAA GCG GCC	913		
35	Lys Val Arg Met Tyr Val Gly Gly Ile Glu His Arg Leu Glu Ala Ala			
	290	295	300	
	TGC AAC TGG ACG CCG GGC GAA CGT TGC GAT CTG GAA GAC AGG GAC AGG	961		
40	Cys Asn Trp Thr Arg Gly Glu Arg Cys Asp Leu Glu Asp Arg Asp Arg			
	305	310	315	320
	TCC GAG CTC AGC CCA TTG CTG CTG TCC ACT ACG CAG TGG CAG GTC CTT	1009		
45	Ser Glu Leu Ser Pro Leu Leu Leu Ser Thr Thr Gln Trp Gln Val Leu			
	325	330	335	
	CCG TGT TCT TTC ACG ACC CTG CCA GCC TTG TCC ACC GGC CTC ATC CAC	1057		
50				
55				

	Pro Cys Ser Phe Thr Thr Leu Pro Ala Leu Ser Thr Gly Leu Ile His	
	340 345 350	
5	CTC CAC CAG AAC ATT GTG GAC GTG CAG TAC TTG TAC GGG GTA GGG TCA	1105
	Leu His Gln Asn Ile Val Asp Val Gln Tyr Leu Tyr Gly Val Gly Ser	
	355 360 365	
10	AGC ATC GCG TCC TGG ACC ATC AAG TGG GAG TAC GTC GTT CTC CTG TTC	1153
	Ser Ile Ala Ser Trp Thr Ile Lys Trp Glu Tyr Val Val Leu Leu Phe	
	370 375 380	
15	CTC CTG CTT GCA GAC GCG CGC GTC TGC TCC TGC TTG TGG ATG ATG TTA	1201
	Leu Leu Leu Ala Asp Ala Arg Val Cys Ser Cys Leu Trp Met Met Leu	
	385 390 395 400	
20	CTC ATA	1207
	Leu Ile	

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(2) INFORMATION FOR SEQ ID NO:10:

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(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 523 base pairs

(B) TYPE: nucleic acid

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(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

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(iv) ANTI-SENSE: No

(vi) ORIGINAL SOURCE:

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(A) ORIGIN: Hepatitis C virus

(B) CLONE: J1(JM)

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

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G ATC CCA CAA GCC ATC TTG GAT ATG ATC GCT GGT GCT CAC TGG GGA GTC 49
 Ile Pro Gln Ala Ile Leu Asp Met Ile Ala Gly Ala His Trp Gly Val
 1 5 10 15
 CTG GCG GGC ATA GCG TAT TTC TCC ATG GTG GGG AAC TGG GCG AAG GTC 97
 Leu Ala Gly Ile Ala Tyr Phe Ser Met Val Gly Asn Trp Ala Lys Val
 10 20 25 30
 CTG GTA GTG CTG TTG CTG TTT GCC GGC GTC GAC GCG GAA ACC ATC GTC 145
 Leu Val Val Leu Leu Leu Phe Ala Gly Val Asp Ala Glu Thr Ile Val
 15 35 40 45
 TCC GGG GGA CAA GCC GCC CGC GCC ATG TCT GGA CTT GTT AGT CTC TTC 193
 Ser Gly Gly Gln Ala Ala Arg Ala Met Ser Gly Leu Val Ser Leu Phe
 20 50 55 60
 ACA CCA GGC GCT AAG CAG AAC ATC CAG CTG ATC AAC ACC AAC GGC AGT 241
 Thr Pro Gly Ala Lys Gln Asn Ile Gln Leu Ile Asn Thr Asn Gly Ser
 25 65 70 75 80
 TGG CAC ATC AAT AGC ACG GCC TTG AAC TGC AAT GAA AGC CTT AAC ACC 289
 Trp His Ile Asn Ser Thr Ala Leu Asn Cys Asn Glu Ser Leu Asn Thr
 30 85 90 95
 GGC TGG TTA GCA GGG CTT ATC TAT CAA CAC AAA TTC AAC TCT TCG GGC 337
 Gly Trp Leu Ala Gly Leu Ile Tyr Gln His Lys Phe Asn Ser Ser Gly
 35 100 105 110
 TGT CCC GAG AGG TTG GCC AGC TGC CGA CGC CTT ACC GAT TTT GAC CAG 385
 Cys Pro Glu Arg Leu Ala Ser Cys Arg Arg Leu Thr Asp Phe Asp Gln
 40 115 120 125
 GGC TGG GGC CCT ATC AGT CAT GCC AAC GGA AGC GGC CCC GAC CAA CGC 433
 Gly Trp Gly Pro Ile Ser His Ala Asn Gly Ser Gly Pro Asp Gln Arg
 45 130 135 140
 CCC TAT TGT TGG CAC TAC CCC CCA AAA CCT TGC GGT ATC GTG CCC GCA 481
 50

Pro Tyr Cys Trp His Tyr Pro Pro Lys Pro Cys Gly Il Val Pro Ala
 145 150 155 160

5 AAG AGC GTA TGT GGC CCG GTA TAT TGC TTC ACT CCC AGC CCC 523

Lys Ser Val Cys Gly Pro Val Tyr Cys Phe Thr Pro Ser Pro
 165 170

10

(2) INFORMATION FOR SEQ ID NO:11:

15

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 523 base pairs

20

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

25

(iv) ANTI-SENSE: No

30

(vi) ORIGINAL SOURCE:

(A) ORIGIN: Hepatitis C virus

(B) CLONE: J4(JM)

35

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

40

G ATC CCA CAA GCT GTC GTG GAC ATG GTG GCG GGG GCC CAC TGG GGA GTC 49

Ile Pro Gln Ala Val Val Asp Met Val Ala Gly Ala His Trp Gly Val

1 5 10 15

45

CTG GCG GGC CTT GCC TAC TAT TCC ATG GTA GGG AAC TGG GCT AAG GTC 97

Leu Ala Gly Leu Ala Tyr Tyr Ser Met Val Gly Asn Trp Ala Lys Val

20 25 30

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CTG ATT GTG GCG CTA CTC TTC GCC GGC GTT GAC GGG GAG ACC TAC ACG 145

55

Leu Ile Val Ala Leu Leu Phe Ala Gly Val Asp Gly Glu Thr Tyr Thr
 35 40 45
 5 TCG GGG GGG GCG GCC AGC CAC ACC ACC TCC ACG CTC GCG TCC CTC TTC 193
 Ser Gly Gly Ala Ala Ser His Thr Thr Ser Thr Leu Ala Ser Leu Phe
 50 55 60
 10 TCA CCT GGG GCG TCT CAG AGA ATC CAG CTT GTG AAT ACC AAC GGC AGC 241
 Ser Pro Gly Ala Ser Gln Arg Ile Gln Leu Val Asn Thr Asn Gly Ser
 65 70 75 80
 15 TGG CAC ATC AAC AGG ACT GCC CTA AAC TGC AAT GAC TCC CTC CAC ACT 289
 Trp His Ile Asn Arg Thr Ala Leu Asn Cys Asn Asp Ser Leu His Thr
 85 90 95
 20 GGG TTC CTT GCC GCG CTG TTC TAC ACA CAC AGG TTC AAC TCG TCC GGG 337
 Gly Phe Leu Ala Ala Leu Phe Tyr Thr His Arg Phe Asn Ser Ser Gly
 100 105 110
 25 TGC CCG GAG CGC ATG GCC AGC TGC CGC CCC ATT GAC TGG TTC GCC CAG 385
 Cys Pro Glu Arg Met Ala Ser Cys Arg Pro Ile Asp Trp Phe Ala Gln
 115 120 125
 30 GGA TGG GGC CCC ATC ACC TAT ACT GAG CCT GAC AGC CCG GAT CAG AGG 433
 Gly Trp Gly Pro Ile Thr Tyr Thr Glu Pro Asp Ser Pro Asp Gln Arg
 130 135 140
 35 CCT TAT TGC TGG CAT TAC GCG CCT CGA CCG TGT GGT ATC GTA CCC GCG 481
 Pro Tyr Cys Trp His Tyr Ala Pro Arg Pro Cys Gly Ile Val Pro Ala
 145 150 155 160
 40 TCG CAG GTG TGT GGT CCA GTG TAT TGC TTC ACC CCA AGC CCT 523
 Ser Gln Val Cys Gly Pro Val Tyr Cys Phe Thr Pro Ser Pro
 165 170
 45
 50
 55

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 402 amino acids

(B) TYPE: protein

(vi) ORIGINAL SOURCE:

(A) ORIGIN: Hepatitis C virus

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Ile Pro Gln Ala Val Val Asp Met Val Ala Gly Ala His Trp Gly Val

Ile Met Ile

Leu

1 5 10 15

Leu Ala Gly Leu Ala Tyr Tyr Ser Met Val Gly Asn Trp Ala Lys Val

Ile Phe Ala

20 25 30

Leu Val Val Met Leu Leu Phe Ala Gly Val Asp Gly Gly Thr His Val

Ile Ala Ala Arg Arg Thr

Leu His Tyr Arg

Asp Ile

Gln Gln

Glu His

Thr Thr

35 40 45

Thr Gly Gly Lys Val Ala Tyr Thr Thr Gln Gly Phe Thr Pro Phe Phe

Ser Val Ala Val Gln Gly His Val Val Ser Arg Leu Val Ser Leu Leu

Met Ala Ala Ser Lys Ser Met Asn Ser Val Ala Arg Met

Arg Ser Ala Ala Thr Thr Ile Gly Trp

Ser Arg Arg His Ala

Gln Phe Gly Leu
 His His Tyr
 Asn Ala
 Ile

50 55 60
 Ser Arg Gly Pro Ser Gln Lys Ile Gln Leu Val Asn Thr Asn Gly Ser
 Arg Pro Ala Gln Glu Arg Val Ile Lys
 Thr Ser Ser Lys Asn
 Ala Gln Ala Asp
 Asn Ala Arg
 Leu

65 70 75 80
 Trp His Ile Asn Arg Thr Ala Leu Asn Cys Asn Asp Ser Leu Asn Thr
 Leu Ser Glu Gln Ala
 Lys
 His

85 90 95
 Gly Phe Leu Ala Ala Leu Phe Tyr Thr His Ser Phe Asn Ala Ser Gly
 Trp Ile Thr Ile Arg Lys Asp Ser Arg
 Gly Ala Arg
 His Gly
 Gln

100 105 110
 Cys Pro Glu Arg Met Ala Gly Cys Arg Pro Ile Asp Glu Phe Ala Gln
 Ala Glu Ser Val Leu Ser Cys Ser Leu Ser Lys Asp
 Gln Gln Thr Thr Trp
 Phe Asp
 Arg Thr

115 120 125

Gly Trp Gly Pro Ile Thr His Val Val Pro Asn Ile Ser Asp Gln Arg
 Asp Ser Tyr Ala Gln Ser Asp Val Pro Glu Glu Lys
 5 Asp Glu Arg Ser Asn Thr
 Thr Met Gly Glu Arg Gly
 Asn Asn Gln Arg Ser
 10 Lys Gly Gly
 Thr
 130 135 140
 15 Pro Tyr Cys Trp His Tyr Ala Pro Arg Pro Cys Gly Ile Val Pro Ala
 Pro Pro Gln Thr Val
 20 Lys
 145 150 155 160
 Ser Gln Val Cys Gly Pro Val Tyr Cys Phe Thr Pro Ser Pro Val Val
 25 Trp Glu
 Lys Ser
 165 170 175
 30 Val Gly Thr Thr Asp Arg Phe Gly Ala Pro Thr Tyr Asn Trp Gly Asn
 Ser Val Thr Ala
 Arg Asp
 35 Ser Glu
 180 185 190
 Asn Glu Thr Asp Val Leu Leu Leu Asn Asn Thr Arg Pro Pro Gln Gly
 40 Asp Phe Val Ser His
 Ile Leu
 195 200 205
 45 Asn Trp Phe Gly Cys Thr Trp Met Asn Gly Thr Gly Phe Thr Lys Thr
 Ser Val
 210 215 220
 50 Cys Gly Gly Pro Pro Cys Asn Ile Gly Gly Val Gly Asn Asn Thr Leu
 55

	Ala		His		Arg		Arg												
			Val				Ala												
5	225		230				235												240
	Thr	Cys	Pro	Thr	Asp	Cys	Phe	Arg	Lys	His	Pro	Glu	Ala	Thr	Tyr	Thr			
	Val											Asp						Ser	
10	His																		
	Leu																		
	Arg																		
15			245				250												255
	Lys	Cys	Gly	Ser	Gly	Pro	Trp	Leu	Thr	Pro	Arg	Cys	Leu	Val	His	Tyr			
	Arg						Ile					Met		Asp					
20			260				265							270					
	Pro	Tyr	Arg	Leu	Trp	His	Tyr	Pro	Cys	Thr	Val	Asn	Phe	Thr	Ile	Phe			
											Ile	Tyr		Val					
25																			Thr
			275				280							285					
30	Lys	Val	Arg	Met	Tyr	Val	Gly	Gly	Val	Glu	His	Arg	Leu	Glu	Ala	Ala			
	Ile								Ile					Asp					
														Asn					
35														Ile					
			290				295							300					
40	Cys	Asn	Trp	Thr	Arg	Gly	Glu	Arg	Cys	Asp	Leu	Glu	Asp	Arg	Asp	Arg			
						Asp			Asn										
	305					310								315					320
45	Ser	Glu	Leu	Ser	Pro	Leu	Leu	Leu	Ser	Thr	Thr	Glu	Trp	Gln	Val	Leu			
	Ala								Thr			Gln		Ile					
	Pro																		
50			325								330								335
	Pro	Cys	Ser	Phe	Thr	Thr	Leu	Pro	Ala	Leu	Ser	Thr	Gly	Leu	Ile	His			

340 345 350
 Leu His Gln Asn Ile Val Asp Val Gln Tyr Leu Tyr Gly Ile Gly Ser
 5 Arg Val Val
 355 360 365
 10 Ala Val Val Ser Phe Ala Ile Lys Trp Glu Tyr Ile Leu Leu Leu Phe
 Ser Ile Ala Ile Val Arg Val Val
 Trp Thr
 15 370 375 380
 Leu Leu Leu Ala Asp Ala Arg Val Cys Ala Cys Leu Trp Met Met Leu
 Ser
 20 385 390 395 400
 Leu Ile

25 (2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

30 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 35 (D) TOPOLOGY: linear

40 (ii) MOLECULAR TYPE: the other nucleic acid (synthesized DNA for
 PCR)

45 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

GCTATCAGCA GCATCATCCA

20

50
 (2) INFORMATION FOR SEQ ID NO:14:

55

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 22 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULAR TYPE: the other nucleic acid (synthesized DNA for PCR)

() SEQUENCE CHARACTERISTIC: N represents inosine.

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

CAGTANTCC GGATCCCNCA AG

22

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 17 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULAR TYPE: the other nucleic acid (synthesized DNA for PCR)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

GTAAAACGAC GGCCAGT

17

5 (2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

10

(A) LENGTH: 17 base pairs

(B) TYPE: nucleic acid

15

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

20

(ii) MOLECULAR TYPE: the other nucleic acid (synthesized DNA for PCR)

25

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

CAGGAAACAG CTATGAC

17

30

(2) INFORMATION FOR SEQ ID NO:17:

35

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 10 base pairs

(B) TYPE: nucleic acid

40

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

45

(ii) MOLECULAR TYPE: the other nucleic acid (synthesized DNA for PCR)

50

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

55

GGACTAGTCC

10

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 16 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULAR TYPE: the other nucleic acid (synthesized DNA for PCR)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

CTAGAGAATT CGGTAC

16

Claims

1. A diagnostic reagent for hepatitis C, which detects an antibody induced by infection of hepatitis C virus, characterized in that said diagnostic reagent comprises the second envelope protein or the first non-structural protein which is encoded by the gene of hepatitis C virus and has a sugar chain.
2. The diagnostic reagent for hepatitis C according to Claim 1, wherein the second envelope protein or the first non-structural protein is represented by an amino acid sequence selected from SEQUENCE ID Nos.1-12 of SEQUENCE LISTING or an amino acid sequence having the same as a part thereof.
3. The diagnostic reagent for hepatitis C according to Claim 1, wherein the second envelope protein or the first non-structural protein is encoded by a base sequence selected from SEQUENCE ID Nos.1-2, and 4-11 of SEQUENCE LISTING.
4. The diagnostic reagent for hepatitis C according to Claim 1, wherein the second envelope protein or the first non-structural protein is produced by an animal cell.
5. The diagnostic reagent for hepatitis C according to Claim 4, wherein the animal cell is CHO cell.
6. A method for detecting an anti-hepatitis C virus antibody, wherein the second envelope protein or the first non-structural protein which is encoded by the gene of hepatitis C virus and has a sugar chain is used as an antigen to detect the antibody specific to said antigen.
7. The method according to Claim 6, wherein the second envelope protein or the first non-structural protein is represented by an amino acid sequence selected from SEQUENCE ID Nos.1-12 of SEQUENCE LISTING or an amino acid sequence having the same as a part thereof.

8. The method according to Claim 6, wherein the second envelope protein or the first non-structural protein is encoded by a base sequence selected from SEQUENCE ID Nos.1-2, and 4-11 of SEQUENCE LISTING.

5 9. The method according to Claim 9, wherein the second envelope protein or the first non-structural protein is produced by an animal cell.

10. The method according to Claim 6, wherein the animal cell is CHO cell.

10 11. A method for detecting an anti-hepatitis C virus antibody, which comprises the steps of contacting a sample with the second envelope protein or the first non-structural protein which is encoded by the gene of hepatitis C virus and has a sugar chain under the conditions that the second envelope protein or the first non-structural protein is bound to the anti-hepatitis C virus antibody to form an immunological complex and measuring the formation of the immunological complex to confirm the
15 presence of the anti-hepatitis C virus antibody in the sample.

12. The method according to Claim 11, wherein the formation of the immunological complex is measured by RIA, ELISA, fluorescent antibody technique, agglutination reaction, or immune precipitation.

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FIG. 1

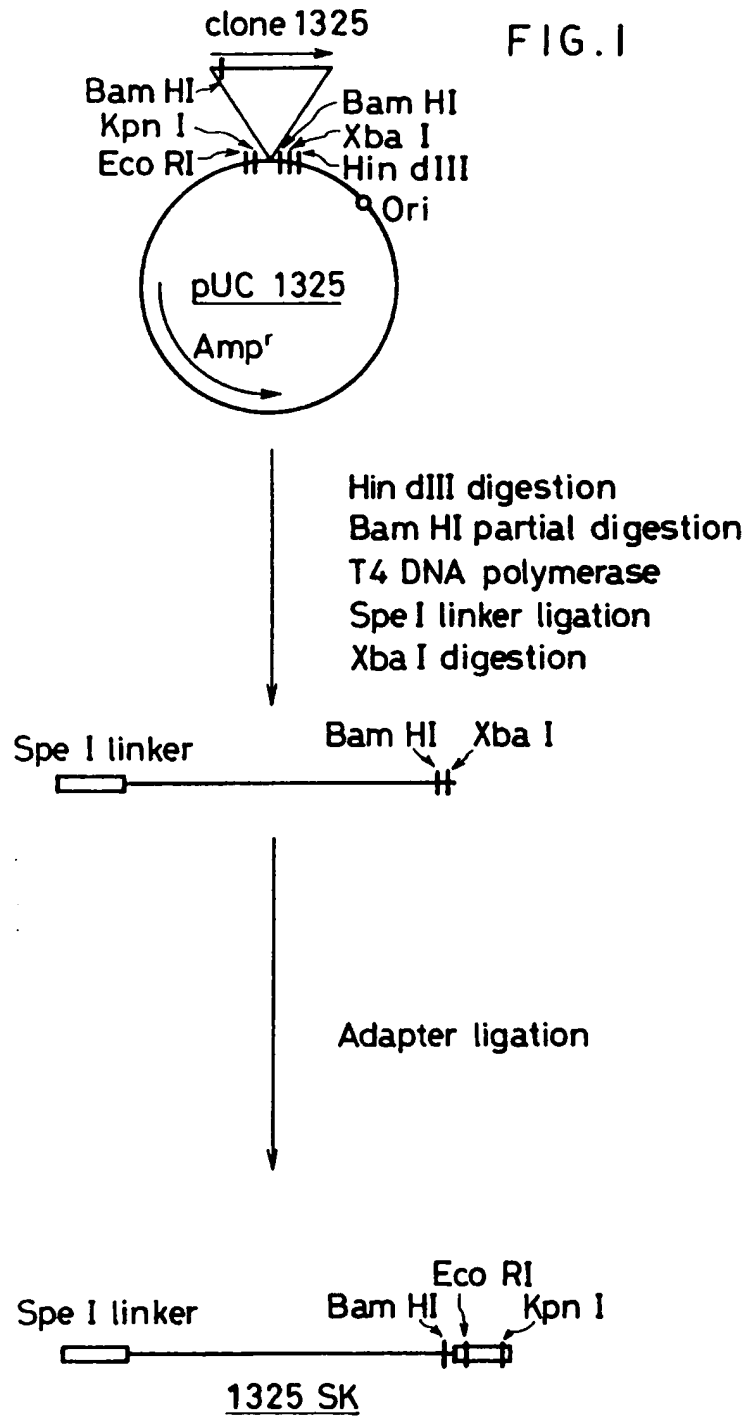


FIG.2

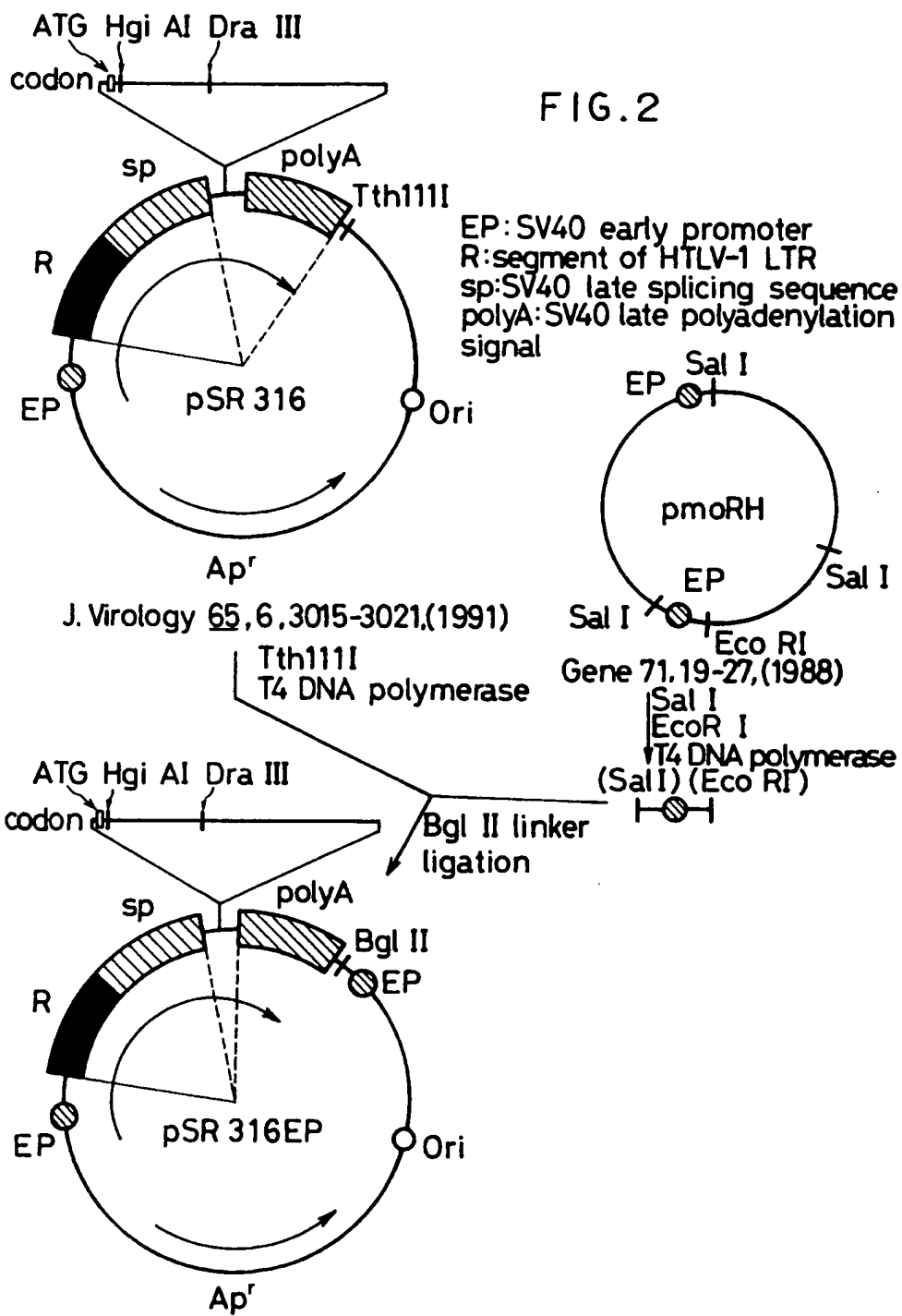


FIG. 3

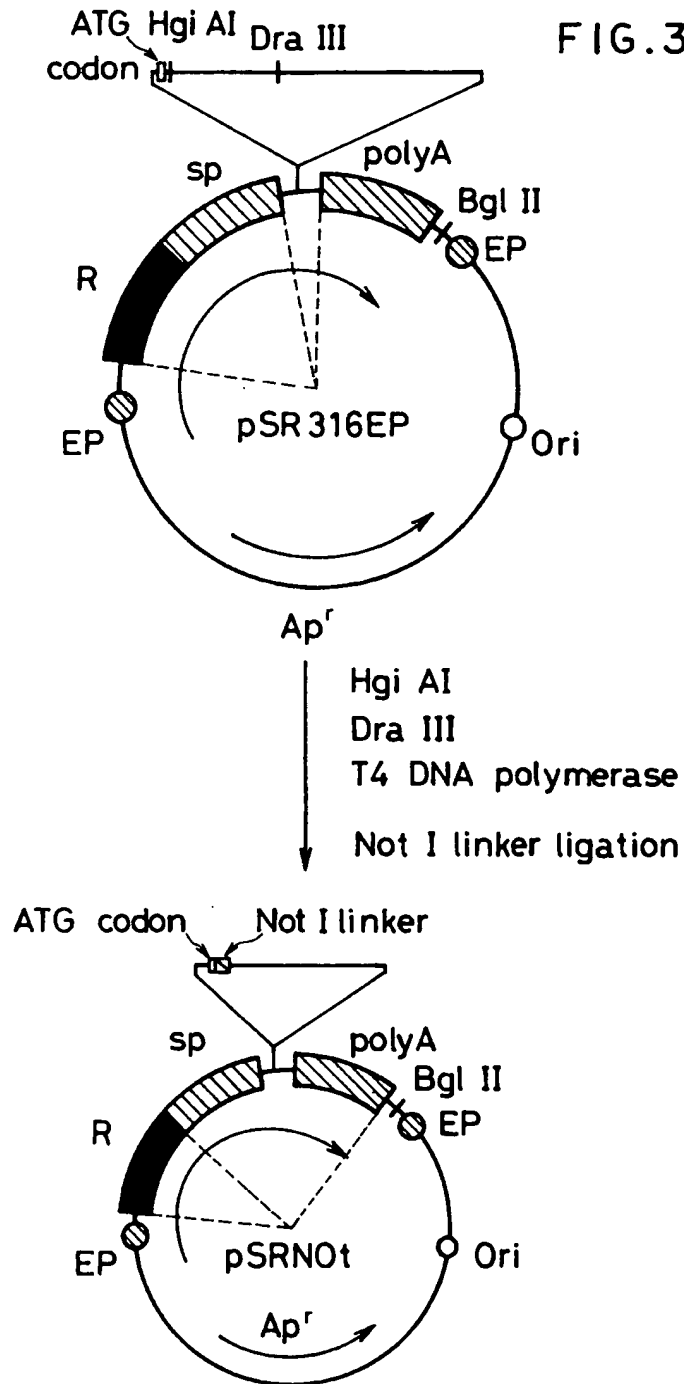


FIG. 4

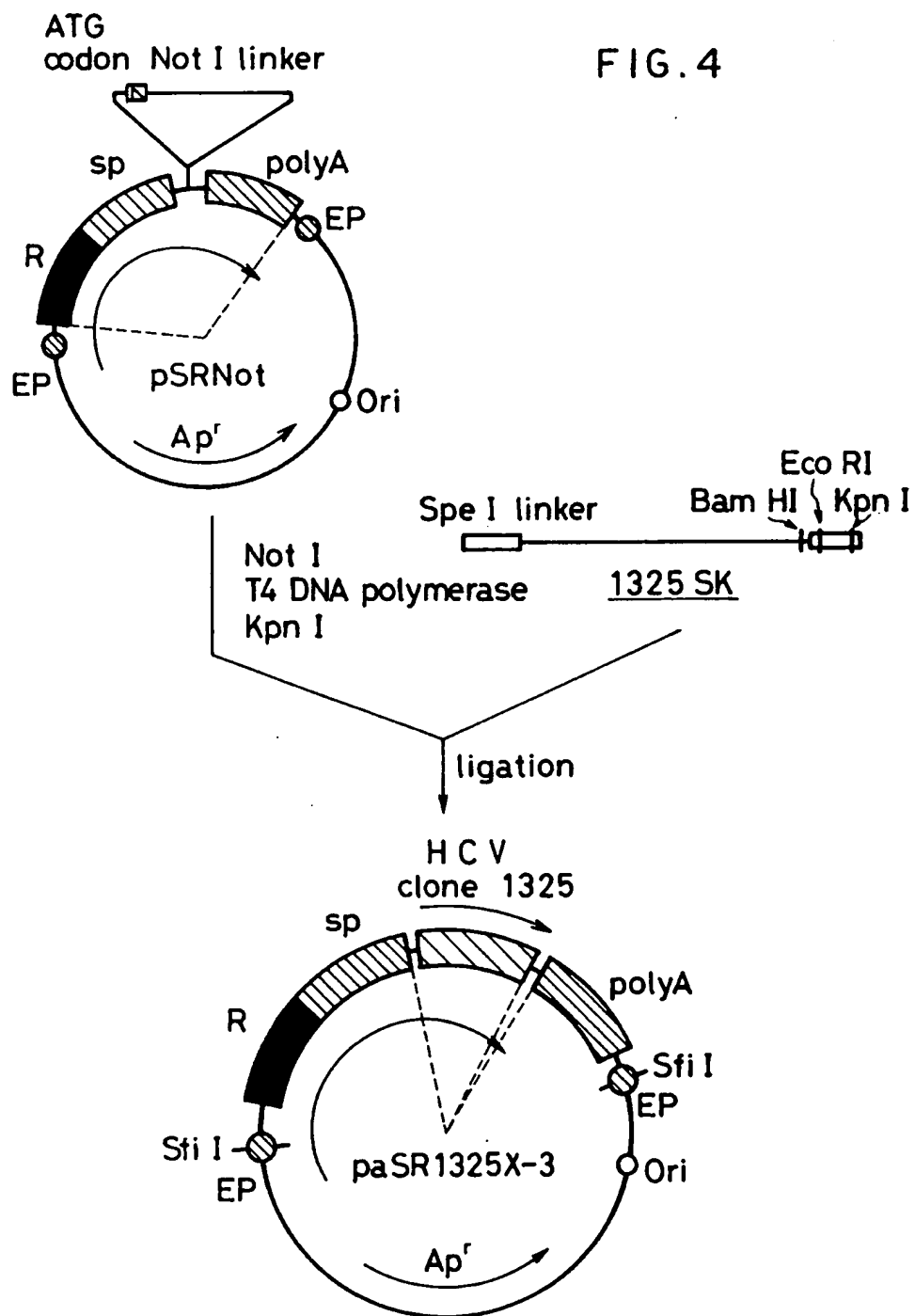


FIG. 5

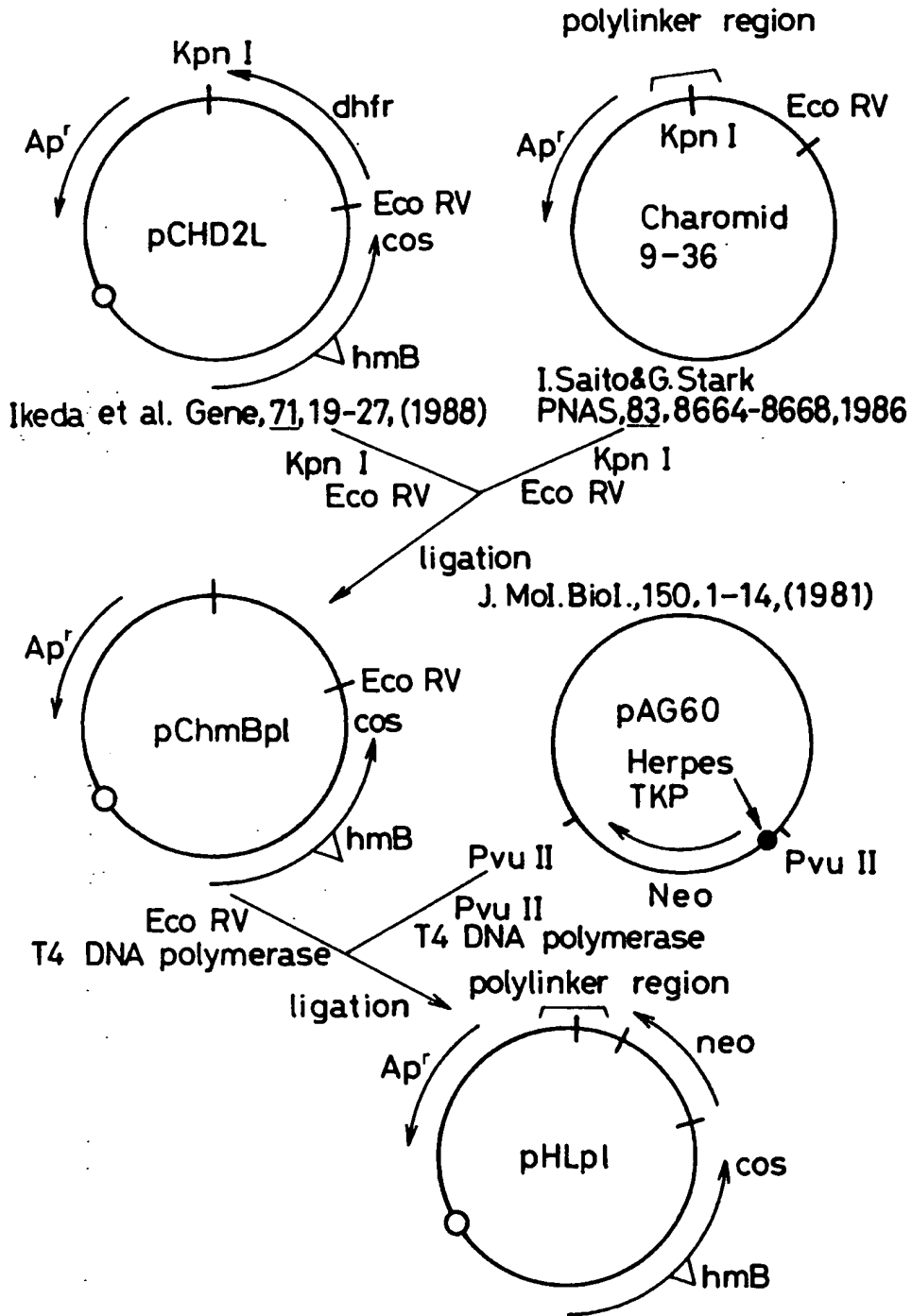
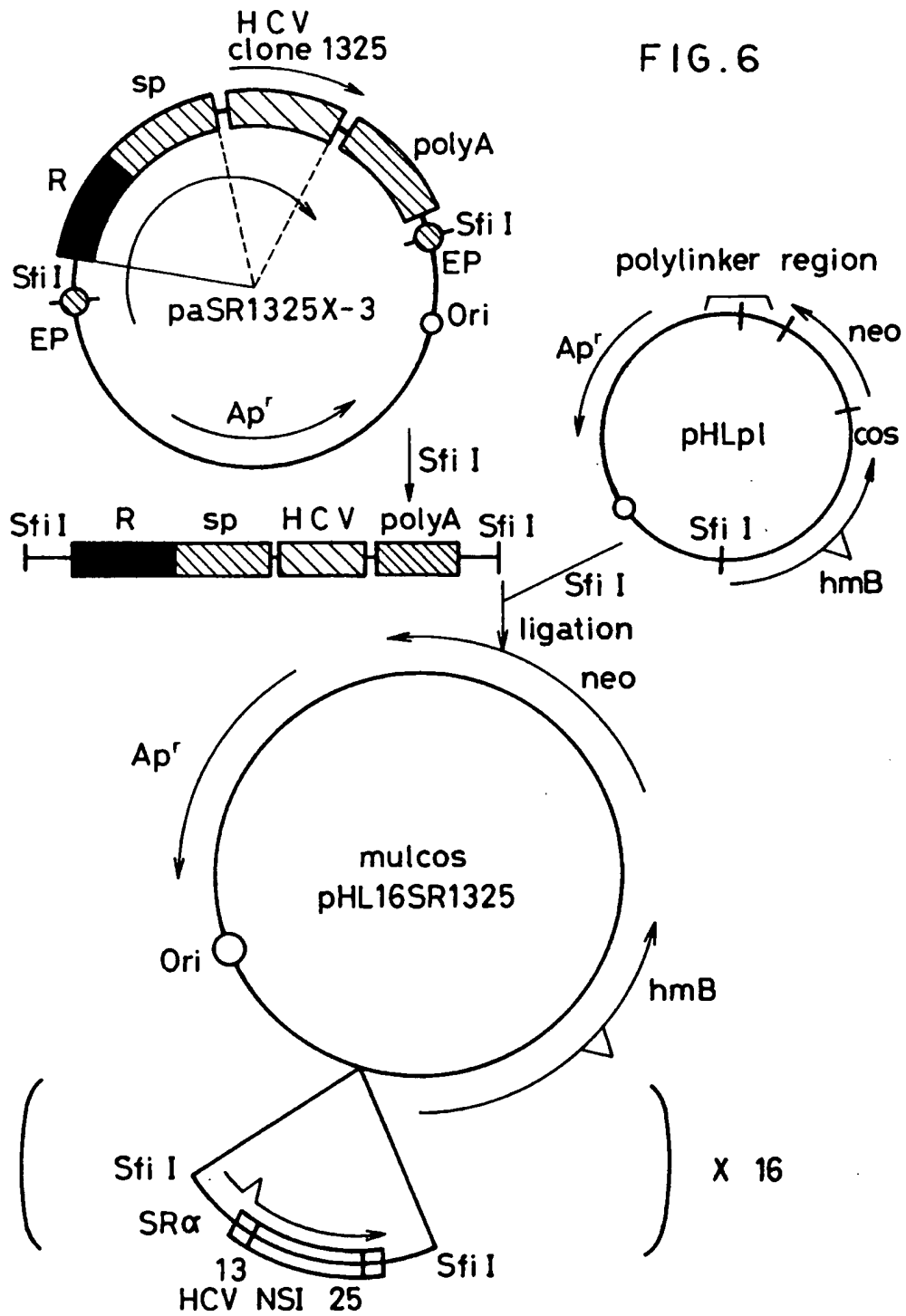


FIG. 6





European Patent
Office

EUROPEAN SEARCH REPORT

Application Number

EP 92 11 7191

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl.5)
X	F. BLAINE HOLLINGER 'VIRAL HEPATITIS AND LIVER DISEASE' 1 June 1991, WILLIAMS & WILKENS, BALTIMORE MD USA See table I in article by G. Kuo et al.: "Serodiagnosis of hepatitis C viral infection using recombinant-based assays for circulating antibodies to different viral proteins." on page 347 - 349. ---	1	G01N33/576 C07K15/00
Y	EP-A-0 388 232 (CHIRON CORPORATION) * page 21, line 9 - line 27 * * page 34, line 6 - line 12 * ---	1-12	
Y	VIROLOGY vol. 180, 1 February 1991, WASHINGTON DC USA pages 842 - 848 A.J. WEINER ET AL. 'Variable and hypervariable domains are found in the regions of HCV corresponding to the Flavivirus envelope and NS1 proteins and the Pestivirus envelope glycoproteins.' * the whole document * ---	1-12	
T	HEPATOLOGY vol. 16, no. 4, 1992, WASHINGTON DC USA page 226A O. YOKOSUKA ET AL. 'Detection of anti-hepatitis c virus E2/NS1 antibody in patients with type c liver disease by western blotting.' * the whole document * -----	1-12	
The present search report has been drawn up for all claims			
Place of search THE HAGUE		Date of completion of the search 19 JANUARY 1993	Examiner VAN BOHEMEN C.G.
CATEGORY OF CITED DOCUMENTS			
X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document		T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons ----- & : member of the same patent family, corresponding document	